



Environmental and socioeconomic risk factors associated with visceral and cutaneous leishmaniasis: a systematic review

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Abstract

We performed a systematic review of the literature published since 1900 about leishmaniasis a neglected vector-borne disease, focused on environmental and social risk factors for visceral (VL) and cutaneous leishmaniasis (CL) to better understand their impact on the incidence of disease. The search terms were “leishmaniasis” AND “risk factors” using Google Scholar, PubMed, and Scielo. We reviewed 177 articles, 95 studies for VL, 75 for CL, and 7 on both forms. We identified 14 categories of risk factors which were divided into three groups: socioeconomic (7), environmental (5), and climate (2) variables. Socioeconomic factors were also associated with disease incidence in vulnerable human populations of arid and tropical developing regions. Environmental and climate factors showed significant associations with the incidence of VL and CL in all the studies that considered them. Proximity to natural vegetation remnants increased disease risk in both the New and Old World while the climate conditions favorable for disease transmission differed among regions. We propose a common conceptual framework for both clinical forms that highlights networks of interaction among risk factors. In both clinical forms, the interplay of these factors played a major role in disease incidence. Although there are similarities in environmental and socioeconomic conditions that mediate the transmission cycle of tropical, arid, and Mediterranean regions, the behavior of vector and reservoirs in each region is different. Special attention should be given to the possibility of vector adaptation to urban environments in developing countries where populations with low socioeconomic status are particularly vulnerable to the disease.

Keywords Leishmaniasis · Risk factors · Socioeconomic · Landscape · Climate

Introduction

Leishmaniasis is a vector-borne disease caused by a protozoan of the genus *Leishmania*, which comprises 20 species

pathogenic to humans (Akhoundi et al. 2016). *Leishmania* is endemic in 97 countries; more than 350 million people are at risk for the disease, and approximately 50,000 to 90,000 cases of VL and 0.7 to 1.2 million cases of CL occur each year. A tentative estimate of mortality based on sparse data using hospital-based fatalities reported 20,000 to 30,000 deaths of leishmaniasis per year in the world (Alvar et al. 2012; World Health Organization 2017). Over 90% of global cases of most fatal leishmaniasis infections are from VL and occur in Brazil, Ethiopia, Sudan, South Sudan, India, and Bangladesh (World Health Organization 2010; Pigott et al. 2014a). In addition, the 84% of global CL incidence in 2016 were reported in Afghanistan, Algeria, Brazil, Colombia, Iraq, Pakistan, Peru, the Syrian Arab Republic, Tunisia, and Yemen (World Health Organization 2018).

The disease has two main clinical forms in humans, visceral (VL) and cutaneous leishmaniasis (CL). In the Old World, VL is caused by parasites of the *Leishmania donovani* species and *L. infantum* in the New World. The cutaneous form, CL, is caused by five species of *Leishmania*: *L. major*, *L. tropica*, *L.*

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aethiopica, *L. donovani*, and in some cases *L. infantum* in the Old World and by multiple phylogenetically distinct *Leishmania* species in the New World (World Health Organization 2010). The clinical manifestations that will develop depend on the *Leishmania* species and host immunological response. Visceral leishmaniasis could present splenomegaly, hepatomegaly, or affect lymphoid tissues. Depending on the form, cutaneous leishmaniasis could present cutaneous nodules, non-ulcerative nodules, or naso-buccal mucosal tissue destruction (Pace 2014).

A total of 93 species of sand flies are probable vectors of *Leishmania* but information on species-specific infection rates is scarce because it is difficult to find infected sand flies in the wild (World Health Organization 2010; Akhoundi et al. 2016). There are two types of *Leishmania* vectors: generalists, which support the growth of more than one species; and specialists, that support only one species of *Leishmania* (World Health Organization 2010). In the Old World, *Phlebotomus* is the principal genus of vectors for the parasite responsible for both VL and CL forms. In the New World, *Lutzomyia longipalpis* is the main vector of *L. infantum*, responsible for VL, while multiple sand fly vectors transmit the various *Leishmania* species responsible for CL (Bates et al. 2015). In the absence of known vectors, however, many sand fly species have been considered as potential *Leishmania* vectors albeit without corroboratory evidence (Akhoundi et al. 2016).

The complex transmission cycle of leishmaniasis also includes several species of mammals other than humans which can be hosts and/or reservoirs of *Leishmania* spp. (Roque and Jansen 2014). Any mammal infected with the parasite can act as a host and may or may not be important in transmission (Roque and Jansen 2014) while reservoirs are only those mammal species highly competent for *Leishmania* spp., responsible for maintaining the parasite in nature (Ashford 1996; Haydon et al. 2002; Roque and Jansen 2014). One important reservoir is the domestic dog (*Canis familiaris*), which is the main reservoir of *L. infantum* in urban areas and is largely responsible for VL around the world and for CL in the Old World. Humans are the reservoir of *Leishmania donovani* responsible for anthroponotic VL transmission in East Africa, Bangladesh, Nepal, Bhutan, and India; and also of *L. tropica* responsible for anthroponotic CL in South-East Asia (World Health Organization 2010). In the New World, other specific reservoirs related to the parasite responsible for CL have not been identified (Reyes and Arrivillaga 2009) and in peri-urban areas close to the forest, other wild mammals are involved in the transmission cycle, especially small mammals such as rodents and marsupials (Roque and Jansen 2014).

The cycle of *Leishmania* spp. depends on a successful transmission between the vector and the host/reservoir. *Leishmania* have two developmental stages: amastigotes inside the macrophages of the mammals and promastigotes in the digestive track of the sand fly (World Health Organization

2010). Sand fly females acquire macrophage infected with amastigotes of *Leishmania* when they feed on blood of a mammal infected. After, the blood meal, amastigotes transform into promastigotes and mature and divide within 3 days of ingestion in the midgut. Then, promastigotes migrate to the proboscis of the sand fly and are ready to be regurgitated into the skin of the vertebrate in the next blood meal (Dawit et al. 2013; Pace 2014; Alemayehu and Alemayehu 2017).

The presence of vector species and potential mammalian reservoirs is favored by some environmental and climatic conditions, such as warm climates and the presence of forest (Desjeux 2001). Such favorable conditions allow sand fly development, providing shelter and protection for both the vector and the reservoir. In fact, both clinical forms of leishmaniasis were initially found only in natural, undisturbed environments (Grimaldi and Tesh 1993). Over the past few decades, however, human migration has led to the creation of settlements close to natural ecosystems, where the cycle of leishmaniasis was already established, increasing human exposure to infected sand flies (Desjeux 2001; Dujardin 2006). These developments have changed the ecology of vectors since the parasite adapted its cycle to peri-domestic sand fly species and reservoir animals (e.g., dogs).

Leishmaniasis transmission occurrence became particularly favorable in developing countries. One potentially important factor underlying this trend is the large human populations of low socioeconomic status living in recently expanded peri-urban areas close to forest or dense vegetation. Peri-urban settlements in these countries are often characterized by poor housing and sanitary conditions that facilitate human contact with vectors, increasing the abundance of potential reservoirs near houses and possibly reducing the efficiency of control programs (Dantas-Torres and Brandão-Filho 2006). Underreporting, deficiency in vector control, and the lack of treatment options in these regions has turned leishmaniasis into one of the most neglected tropical diseases (Desjeux 2001). In addition, shifts in temperature and precipitation in these developing regions could lead to further changes in vector-borne disease incidence as climate conditions for the sand flies and reservoirs are altered (Moo Llanes 2016).

Previous reviews on the risk factors related to the transmission of the disease focused on visceral leishmaniasis in Asia (Bern et al. 2010; Singh and Singh 2019), Africa (Assefa 2018), and in the Americas (Belo et al. 2013). Additionally, the last reviews describing both clinical forms and their risk factors around the world took place over 10 years ago and did not provide a description of the network of risk factors for each clinical form (Desjeux 2001, 2004; Shaw 2007). As result, how interactions between environmental and socioeconomic factors influence leishmaniasis risk is not well understood. The aims of the present study are to (1) review literature on environmental and social factors associated with leishmaniasis in humans and (2) determine the influence of social,

environment and climate factors on disease incidence in order to make a conceptual model about the dynamic of leishmaniasis transmission. We hope that this approach to data synthesis will help to better understand and highlight the interplay of the network of risk factors that influence the incidence of both clinical forms and to guide control efforts.

Methods

We searched the literature for relevant publications between 1900 and September 2019 using Web of Science (“all fields”), Google Scholar (“with all of the words”), PubMed (“all fields”), and Scielo (“all indexes”). Search terms included both “leishmaniasis” AND “risk factors” together. We examined the titles and abstracts of all articles identified in the searches and the full texts if necessary in order to identify risk factors included in the studies. We did not analyze articles that focused on (1) canine leishmaniasis exclusively; (2) treatment or clinical factors; (3) asymptomatic leishmaniasis (because we cannot identify the factors that influence the occurrence of leishmaniasis); (4) genetics of the disease or its vectors; and (5) leishmaniasis associations with other disease or clinical descriptions. Additional articles were located through citations from the selected articles or from suggestions from disease experts.

All potential articles were screened for risk variables considered in this review. These variables were then split into three categories: socioeconomic and demographic factors, landscape and environment factors, and climatic factors. We identified 14 variables that were most frequently included in the studies: 7 socioeconomic and demographic factors, 5 landscape and environmental factors, and 2 climatic factors (Table 1).

We considered all the regions affected by leishmaniasis around the world to see similarities in disease dynamics for both clinical forms and highlight the differences between Mediterranean regions and tropical and arid regions, where the transmission cycle is similar but the behavior of vector and reservoirs differs.

Results

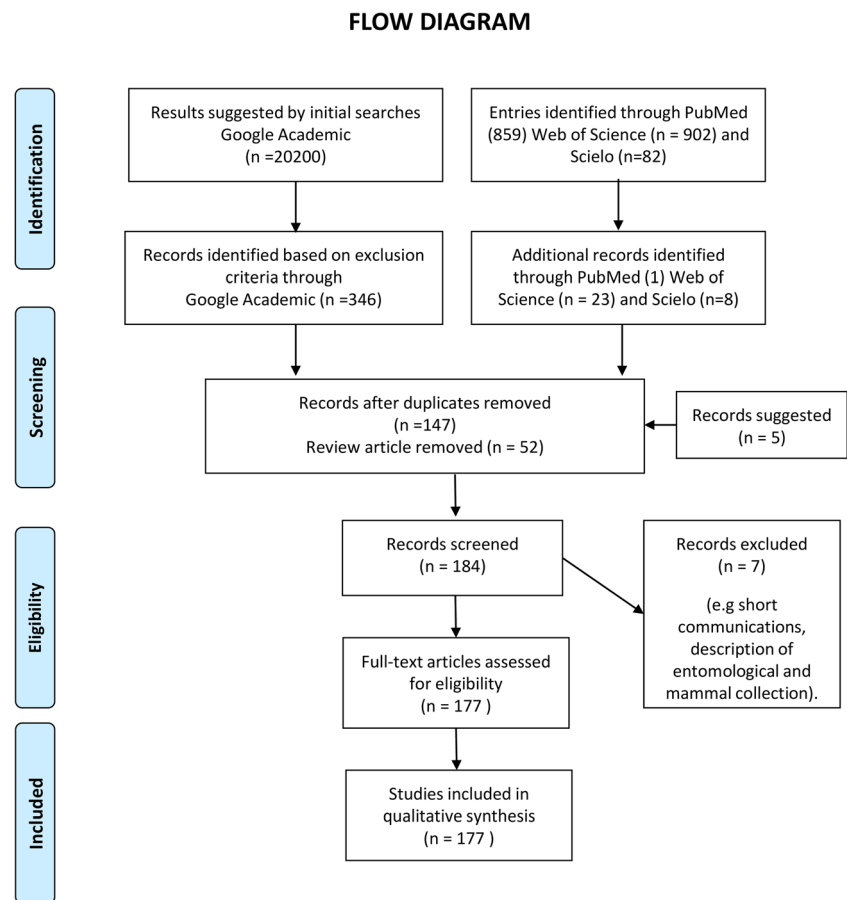
Selection of publications and general description

Our search resulted in 184 potentially relevant articles, of which 177 met our inclusion criteria. Among these 177 articles, 75 focused on cutaneous leishmaniasis (CL), 95 on visceral leishmaniasis (VL), and 7 on both clinical forms (Fig. 1). A total of 52 articles were studies conducted in Brazil, 18 were conducted in Iran, and 15 in Ethiopia, with the remainder distributed across other countries (Fig. 2).

Table 1 List of risk factor variables for leishmaniasis identified in the studies included in the review

Category	Risk variables	Influences
Social and demographic	Socioeconomic status	Influence of quality of life (type of house). Houses built with straw and mud, provide shelter to sand flies.
	Water supply/sewage system/garbage collection	Trash, sewage water, and wells create environmental conditions suitable for sand fly breeding sites.
	Characteristics of population (age, gender, education level, migrant)	Children and elderly people are more vulnerable. Agricultural workers are usually males. Migration increases the number of informal settlements. Low education level can influence the lack of adequate preventive measures.
	Presence of domestic or wild animals	Potential reservoirs or blood meal source.
	Behavior	Sleeping outside without protection against sand flies and working in vegetated areas increase exposure to sand flies.
	Health factors	Immunosuppressed people with other illness or poor nutrition may be vulnerable. Contact with other leishmaniasis cases (anthroponotic transmission).
Landscape and environmental variables	Population density	Household size increase attractiveness to sand flies.
	Vegetation	Vegetation provides shelter to vector and reservoirs.
	Presence of waterbodies	Humid conditions foster sand fly breeding.
	Altitude/slope/soil type	Physical conditions favorable for breeding sites of vectors.
	Urban/rural landscape	Leishmaniasis cycle presence near rural and urban areas, depending on the parasite species, increases the risk.
Climatic	Construction	Anthropogenic disturbance in forested areas increase human-vector contact.
	Temperature/precipitation	Conditions may be favorable to sand fly development.

Fig. 1. Flow diagram of literature search process



For each clinical form, we examined the relationship between the number of times risk variables were included in studies and their significance (Fig. 3). Landscape variables were implicated with incidence of VL in 48% of the studies and in 62% for CL. Climatic variables were associated with VL in 25% of the studies and in 30% for CL. Socioeconomic and demographic variables were most commonly considered for VL and CL in 67% and 63% respectively (Fig. 4).

To summarize the information of the risk factors considered in our review, we extracted metrics of variable significance directly from the original articles. In addition, for the studies (20% of 177) that provided data for odds ratios (OR), the ratio of disease odds given exposure status, or relative risk (RR), the probability of developing leishmaniasis in an exposed group compared with non-exposed group, we used the OR/RR values. For the studies which provide land cover (10%), data were collected using *in situ* observations or satellite images coupled with geographic information systems (GIS). Metrics of vegetation cover near leishmaniasis cases (36%) included the Normalized Difference Vegetation Index (NDVI), classification of the type and extent of vegetation cover within a given radius of the household, and the distance from the household to vegetation. Detailed information for all studies is included in online resource (Table S1).

Socioeconomic variables

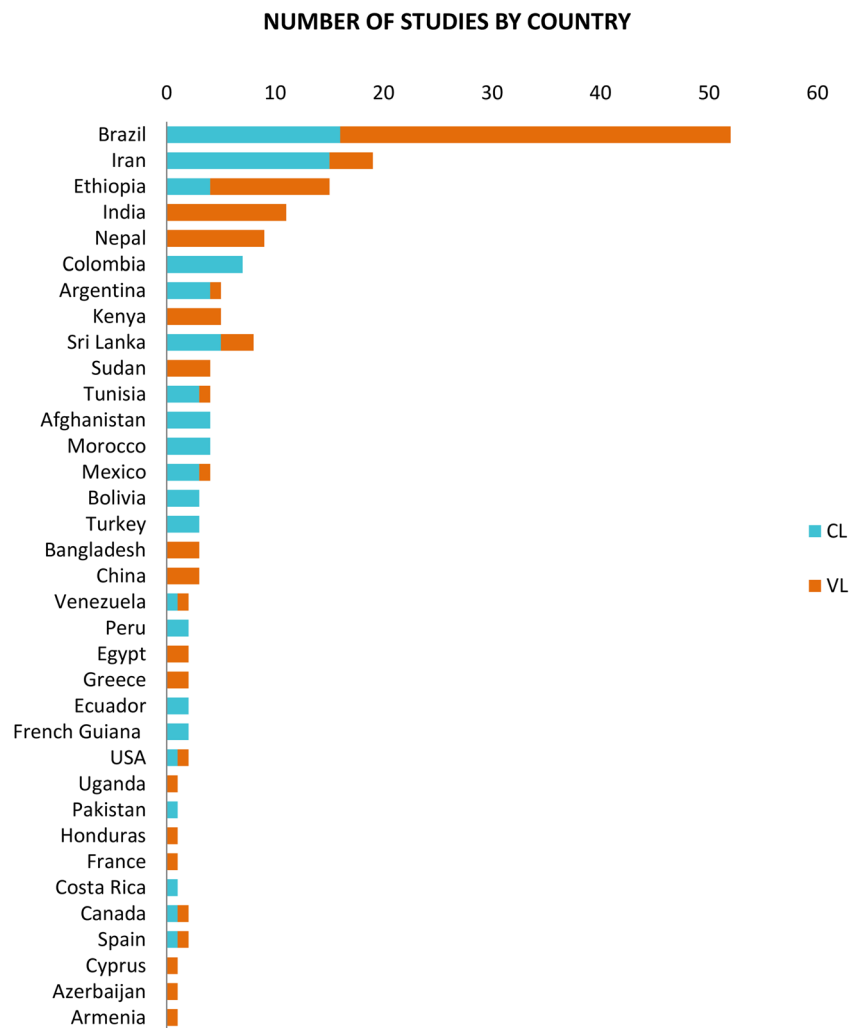
Socioeconomic and demographic factors (Table 1) were significant in 41% of VL and 49% of CL of the cases in the studies (Fig. 3).

Visceral leishmaniasis

Household characteristics that reflect precarious living conditions were analyzed in 44 studies, with 52% finding significant results. For instance, in Posadas, Argentina, the majority of people with VL lived in poor quality houses with sand floors, wooden walls, totally or partially open roofs, and without window screens (López et al. 2016). For anthroponotic VL in Bihar, India mud-plastered walls resulted in a twofold risk increase (OR = 2.40) compared with cement-plastered ones (Ranjan et al. 2005) and in Fulbaria, Bangladesh the prevalence of cases was 59% in houses made of mud floor and tin wall (Bhowmick and Khanum 2017).

Indicators of low educational status were also significant VL risk factors for 39% of the 31 studies that considered this factor. In Belo Horizonte, Minas Gerais, Brazil, a study found a higher risk of VL for illiterate people (RR = 2.87) and for households with less than 4 years of education (RR = 1.82) (de

Fig. 2 Geographic distribution of number of studies on leishmaniasis risk factors examined in this review. (VL visceral, CL cutaneous)



Araújo et al. 2013). For the study in Bihar, India, with anthroponotic VL risk for illiterate people was higher (OR = 1.66) than for literate people (Ranjan et al. 2005).

Lack of access to sewage services, water supply, and garbage collection also presented a significant risk for 33% of the 51 studies that included these factors. In two studies in Brazil, high incidence of VL was associated with lack of these basic sanitary services (Moreno et al. 2005; de Almeida et al. 2011). In these same localities, people with low income were also concentrated in high-risk areas in peripheral neighborhoods for VL (Werneck et al. 2007; de Araújo et al. 2013). A rapid recent increase in population density was also a characteristic of areas of high VL incidence (Cerbino Neto et al. 2009). However, low income in itself was only significant in 34% of 35 studies.

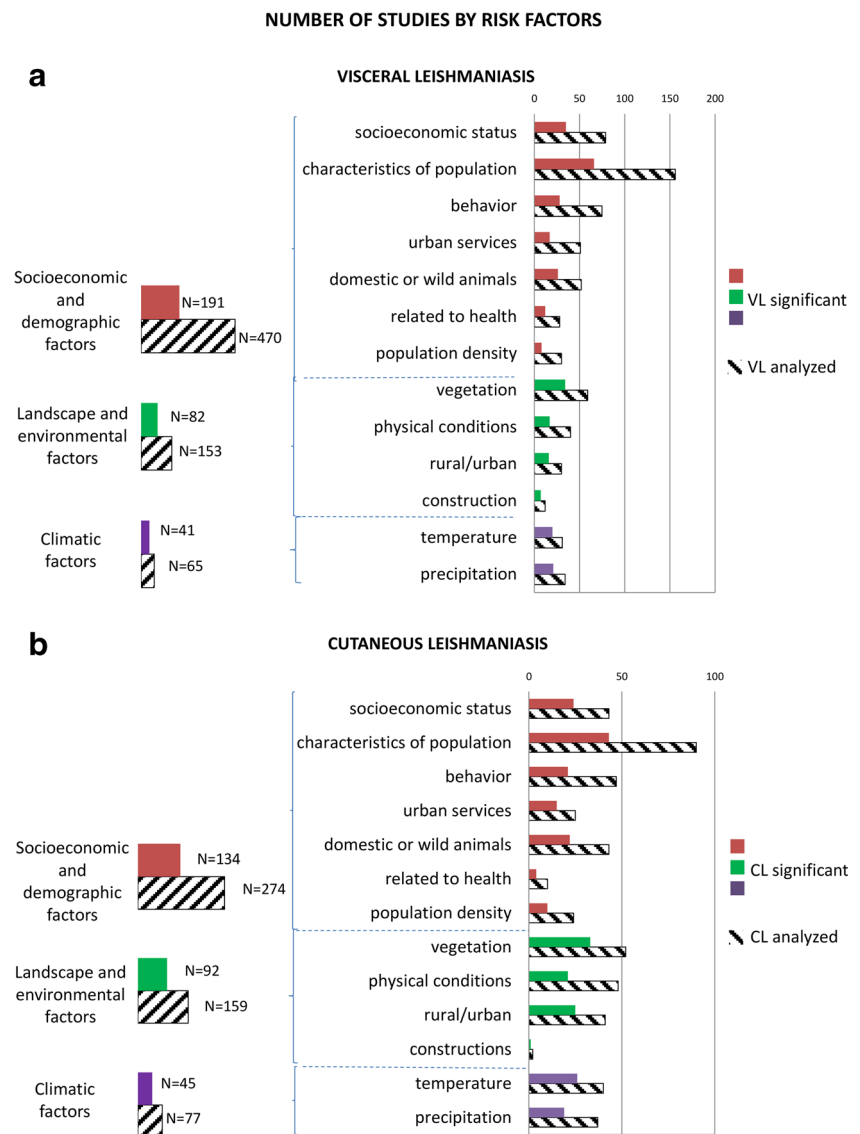
The dog as the main reservoir of *Leishmania infantum* in urban areas is associated with high risks of infection for VL in 67% of 15 studies. For instance, in two studies in Belo Horizonte (RR = 19.11, OR = 2.17) (Borges et al. 2009; de Araújo et al. 2013), and in Posadas where the presence of at least one infected dog was associated with the risk of VL

infection (OR = 120.30) and reported in 100% ($n = 24$) of VL cases analyzed (López et al. 2016).

The presence of other peri-domestic animals was a significant VL risk factor in 47% of 30 studies. In Belo Horizonte, Brazil, the presence of chickens and ducks led to a twofold increase in risk of VL (OR = 2.10) (Moreno et al. 2005). In the same city, another study found a risk increment for people who kept ducks, chickens, and other birds (OR = 4.18, 1.57, 1.47 respectively) (Borges et al. 2009). Presence of rodents (OR = 1.81) in Brazil (Borges et al. 2009) and goats (OR = 6.50) in Tigray, Ethiopia (Yared et al. 2014) also incremented the risk of VL.

Although not strictly socioeconomic conditions, certain activities such as sleeping outside the house, either on the ground (OR = 4.53), under vegetation (OR = 2.77), or near dogs (OR = 4.30) increased the risk ratio of VL in Tigray, Ethiopia (Argaw et al. 2013). Time spent outdoors was also a risk factor for VL in Posadas, Argentina (OR = 4.50) (López et al. 2016) and in Belo Horizonte, Brazil (OR = 1.90) (Moreno et al. 2005). The proximity to the household of a VL patient increase the risk in countries with anthroponotic

Fig. 3 Number of times that each of the three category of risk variables listed in Table 1 were considered in the 176 studies and number reporting significant values. For **a** visceral (VL) and **b** cutaneous (CL) leishmaniasis studies. Left panels show results aggregated for each category. Right panels show subdivisions for each category based on Table 1



transmission as India (OR = 76, RR = 11.89) (Barnett et al. 2005; Perry et al. 2013), Bangladesh (OR = 25.40) (Bern et al. 2005), and Nepal (OR = 3.49) (Mandal et al. 2019). History of another disease in the previous year (OR = 2.76) had a significant impact on the occurrence of VL in Bihar, India (Ranjan et al. 2005). Malnutrition was another risk factor related to health associated with an increase of VL incidence in Jacobina, Bahia, Brazil (Badaró 1988).

Cutaneous leishmaniasis

Among CL studies, household characteristics were significant for 50% of 28 studies. In Salta, Argentina, windows that cannot be locked represented a high risk for CL transmission (OR = 2.93) (Sosa-Estani et al. 2001). Households built with non-durable wall materials had increasing risk of acquiring CL (OR = 2.36) in Alagoas State, Brazil (de Araújo and de Alencar

Ximenes 2009; de Oliveira et al. 2012). In Matara, Sri Lanka, unplastered walls were associated with higher CL risk (Kariyawasam et al. 2015), and in Tigray Ethiopia, the presence of cracks or holes in the walls led to a fourfold risk increase (OR = 4.04) for VL (Bsrat et al. 2015). Household characteristics were also risk factors for anthroponotic CL in Kabul, Afghanistan where brick wall type led to a twofold risk (OR = 2.33) (Reithinger et al. 2010) and poor interior housing conditions (OR = 1.99) in Kerman, Iran (Bamorovat et al. 2018).

Low economic level and related factors are important risk factors for CL (67% of 15 studies). Poverty had a significant association with infection risk in Isfahan, Iran, (OR = 2.03) (Nilforoushzadeh et al. 2014) and in Matara, Sri Lanka (OR = 28.66) (Kariyawasam et al. 2015). Furthermore, the absence of a gas stove in Alagoas (OR = 2.41), in houses of people with low-income status in Brazil (de Araújo and de Alencar Ximenes 2009), were also associated with populations affected with CL.

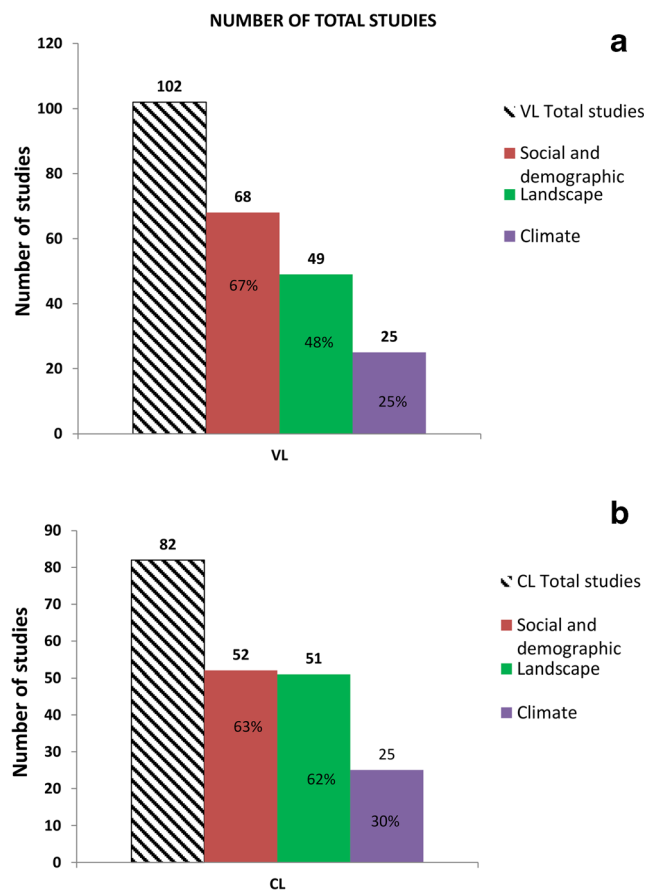


Fig. 4 Number of total studies analyzed. For **a** visceral (VL) and **b** cutaneous leishmaniasis (CL). Hatched bars show total number of studies. Colored bars show the number of studies that found factors in each category significant. Note that some studies examined several factors

As for VL, the presence of peri-domestic animals was important for the transmission of CL (38% of 16 studies), whether they were inside (OR = 2.93) (de Araújo and de Alencar Ximenes 2009) or outside the house (OR = 2.38) (Sosa-Estani et al. 2001; Nilforoushzadeh et al. 2014).

Activities such as sleeping at the workplace (rural work OR = 4.14) and sleeping outdoor at night (OR = 6.29) were associated with higher incidence in Salta, Argentina, (Sosa-Estani et al. 2001). The absence of protection measures against sand flies bites (OR = 6.13) during the time spent outdoors represented a higher risk for CL transmission in Matara, Sri Lanka (OR = 24.60) (Kariyawasam et al. 2015). Agricultural work (OR = 7.75) and leisure activities inside the forest (OR = 9.23) were associated with the incidence of CL in Alagoas State, Brazil (de Araújo and de Alencar Ximenes 2009).

Environmental and landscape factors

These factors are a range of descriptors of the area in which leishmaniasis occurred (Table 1). Land cover data, vegetation

cover near leishmaniasis cases and the distance from the household to vegetation. Also were considered physical variables such as soil type and elevation in a number of studies. These environmental and landscape factors were significant in 54% of VL and 58% of CL of the cases (Fig. 3).

Visceral leishmaniasis

For VL, distance to vegetation was a significant risk factor for 54% of the 26 studies that included this variable. In Bihar, India, the presence of bamboo near the house resulted in a twofold risk increment (OR = 2.30) relative to areas with creepers, herbs, and bushes (Ranjan et al. 2005) and in Dharan, Nepal were proximity to a forest island increase the risk of VL with anthroponotic transmission (Khanal et al. 2010). Similarly, in Teresina, Brazil, the predominant vegetation in the city was shrubs, palm trees, and sparse mango trees. However, Teresina is surrounded by Brazilian Cerrado (Savannah) and pastureland and the high incidence rates were found in the peripheral neighborhoods close to both vegetation types (Werneck et al. 2002).

Vegetation cover type was significant in 60% of the ten studies that considered it, particularly when vegetation was interspersed with urban development in peri-urban areas. In the Gangetic plain of NE India, the presence of woodland (< 10% tree cover) was associated with high anthroponotic VL risk due to the proximity to peri-urban area (Bhunia et al. 2010b). On the other hand, in the Mediterranean city of Fuenlabrada, Spain, a spatial analysis located a cluster of high incidence of leishmaniasis (VL and CL) close to the forest of Bosquesur Park, an urban ecological corridor (Gomez-Barroso et al. 2015). In the Provence-Alpes-Cotes d'Azur, France, a cluster of VL was located in Nice in scattered dwellings close to the Mediterranean mixed forest. These examples demonstrate that this effect is not restricted to leishmaniasis of tropical areas (Faucher et al. 2012).

Fourteen studies considered NDVI index as a predictor of incidence and 10 found significant associations. High NDVI values were associated with incidence of VL in Teresina and Campo Grande with a vegetation typical of Brazilian Cerrado (Cerbino Neto et al. 2009; de Oliveira et al. 2012). However, incidence was higher in areas of low NDVI in Distrito Sanitário de Barra, Bahia, Brazil, where Caatinga (Brazilian Thorny scrub) seasonal loss of leaves in this habitat is associated with low NDVI (Bavia et al. 2005). In India, low NDVI values were also significant for anthroponotic VL incidence because most of the cases occurred in areas with low density of vegetation where live the vector *Phlebotomus argentipes* (Bhunia et al. 2010b, 2012).

Others variables related to land use and geography include the presence of waterbodies near dwellings and the location of study sites in relation to urban or rural areas. The presence of waterbodies close to the dwellings was a risk factor for VL in

6 out of 16 studies. High susceptibility areas of VL were close to the rivers in Kalaybar and Ahar, Iran (Rajabi et al. 2014) and in Sri Lanka, water reservoirs represented also a risk of VL incidence (Weerakoon et al. 2018). In addition, increased risk of anthroponotic VL was greater closer to ponds in Dulari, Dharan, Nepal (OR = 3.70) (Schenkel et al. 2006), rivers and waterbodies in the Gangetic plain, especially in non-perennial river banks in India (Bhunia et al. 2010b, 2011).

The majority of studies of VL risk factors were conducted in urban areas. However, 17 studies specifically consider urban cover as a risk factor with 59% studies finding a significant relationship between urban cover and VL risk. High incidence of VL was centered in peripheral neighborhoods in Teresina, Brazil (Werneck et al. 2002) and in Kyrenia, Cyprus (Ruh et al. 2017). Continuous urban area in Marseille, France (Faucher et al. 2012) and Green urban areas in Attica, Greece (Iliopoulou et al. 2018) presented also a high incidence of VL. In addition, built-up areas in northwest India represented a high anthroponotic VL risk (Bhunia et al. 2010b). However, other studies found high VL risk in rural areas (four of seven studies), for example, cultivated and irrigated land in Thessaly, Greece (Giannakopoulos et al. 2016); and in north-western Iran (Rajabi et al. 2016b). In addition, construction projects such as highways and pipelines were also associated with an increase in VL incidence in Brazil (58% of 12 studies) (Antoniali et al. 2007; Cardim et al. 2013, 2016).

Eight studies considered the effects of soil type on VL and four of these found significant associations. Soil characteristics are hypothesized to reflect moisture conditions suitable for breeding sites; soil types associated with high VL incidence areas include fluvisols in the Gangetic plain in India (anthroponotic transmission), alluvial soil characteristic of the rivers areas as the Ganges river (Bhunia et al. 2010b), and vertisols (OR = 24.32), lixisols, cambisols, and luvisols characteristic of tropical grasslands and savannas in Ethiopia (Tsegaw et al. 2013; Kirstein et al. 2018).

Altitude of the study area has also been associated with VL incidence with 7 of 12 studies finding this variable a significant risk factor. For instance, 95% of VL cases in Ethiopia were located in areas lower than 1872 m asl in a range between 1000 and 3000 m asl (Tsegaw et al. 2013). In Gedaref, Sudan, high risk areas were found in areas lower than 550 m asl (range 400–1000 m asl) (Elnaiem et al. 2003b) and in Thessaly, Greece in areas lower than 200 m asl (range 27–1083 m asl) (Giannakopoulos et al. 2016).

Cutaneous leishmaniasis

Environmental and landscape variables were also high risk factors for CL transmission. Vegetation close to the dwellings was a significant risk factor for 63% of 19 studies. Areas with a predominance of pastures and secondary vegetation close to households in Seropédica, Rio de Janeiro were favorable for

CL occurrence (de Oliveira et al. 2016). In Alagoas State, Brazil, originally covered by Caatinga and Atlantic coastal forest, the presence of forest less than 200 m from households resulted in a fourfold greater risk (OR = 4.70) relative to areas farther from the forest (de Araújo and de Alencar Ximenes 2009). Areas characterized by residual forests and riparian forests resulting from reforestation in Campinas, São Paulo State, Brazil, accounted for 82% of the cases east of the city and 50% of the cases in the southeast occurred in sites less than 200 m from forest (Nasser et al. 2009).

Vegetation cover was also a significant risk factor for 92% of 13 studies which analyzed it for CL. In the district of Matara in Southern Sri Lanka, a spatial analysis found that clusters of CL were more prevalent in proximity to native xeric shrublands (Kariyawasam et al. 2015). CL transmission was also observed in the Colombian Andean region, an area characterized by a mosaic of savanna, rainforest, and woodlands. Among all categories of land cover analyzed in this study, rainforest cover was positively associated with CL incidence (Pérez-Flórez et al. 2016). Other studies conducted in a department located in the same Andean region of Colombia found that areas of high incidence had a 20% higher cover of wooded and shrubs relative to disease free areas (Valderrama-Ardila et al. 2010; Ocampo et al. 2012).

NDVI values had significant associations with CL in 55% of nine studies. In Itapira, Brazil, high NDVI values represented 50% of CL risk areas (Aparicio and Bitencourt 2004) and also CL incidence was associated with NDVI values in Adana, Turkey (Artun and Kavur 2017). In Iran, high NDVI values were significant for CL cases (Ramezankhani et al. 2017b; Shiravand et al. 2018) and with prevalence of anthroponotic CL (Golpayegani et al. 2018).

Other landscape characteristics, such as the extent agricultural area, were associated with CL for 71% of 17 studies. The study areas previously mentioned associated with vegetation were rural areas in the case of Seropédica (de Oliveira et al. 2016) and cultivated and livestock areas in the Colombian Andean region (Pérez-Flórez et al. 2016). Likewise, in Caratinga, Minas Gerais, Brazil, 77% of CL cases were located in rural areas (Machado-Coelho et al. 1999), and in Tsaedemba, Tigray, Ethiopia farm land within 300 m radius from the households (OR = 1.86) increased CL risk (Bsarat et al. 2015). In western Afghanistan, irrigated cultivated lands (OR = 14.34) were the land cover more significantly associated with anthroponotic CL (Fakhar et al. 2017).

Six of 12 studies analyzed distance of households to waterbodies. Population centers affected with CL were characterized by the presence of riverbeds and embankments in Isfahan, Iran (Nilforoushzadeh et al. 2014; Rajabi et al. 2016a) and presence of ponds or waterways in Loreto, Santiago del Estero province in northern Argentina (Yadon et al. 2003). In western Afghanistan, Harirud river ($p < 0.01$) was a risk factor of CL with anthroponotic transmission (Fakhar et al. 2017).

Only four studies of five studies found significant associations of CL with soil type. In Seropédica, Rio de Janeiro, planosol characterized favorable areas of CL (de Oliveira et al. 2016), inceptisol soil type were significant for CL incidence in Ilam and Dehloran, Iran (Mokhtari et al. 2016; Nikonahad et al. 2017). For anthroponotic CL, haplocalcids with torriorthents and torrifluvents soil types had highest association (OR = 13.08) in western Afghanistan (Fakhar et al. 2017).

Altitude and elevation data (slope) were also found to be significant in 11 out of 31 CL studies. In Seropédica (range 0–520 m asl), favorable areas of CL incidence were characterized by low altitude in areas ranging from 0 to 40 m asl and slope of 0–2.5° (de Oliveira et al. 2016). Human contact risk zones (between the vector and human) in Itapira, São Paulo State, Brazil (1–1200 m asl) were at altitudes lower than 750 m asl (Aparicio and Bitencourt 2004). In contrast, altitudes between 1400 and 2700 m asl (OR = 2.32) and slopes higher than 4.6° (OR = 4.36) were most favorable for CL incidence in Ethiopia (range 1000–3000 m asl) (Seid et al. 2014).

Climate variables

Climate variables were analyzed in studies that considered other environmental variables (i.e., vegetation, soil type, altitude). These variables were significant in 63% of VL and 58% of CL of the cases in the studies (Fig. 3). The most commonly studied climate variables for both clinical forms were temperature and rainfall. The approach of most studies was to examine the range of temperatures and rainfall that were favorable to disease incidence. Other studies used Environmental Niche Models to examine the association between climate variables and disease occurrence.

Visceral leishmaniasis

Temperature was significant for 60% of 25 studies, rainfall for 60% of 25 studies, and relative humidity for 67% of 9 studies. Incidence areas in the Gangetic plain, India (anthroponotic transmission) had a temperature range between 25° and 27 °C and precipitation between 100 and 160 mm with relative humidity between 66 and 75% (Bhunia et al. 2010b). In China, temperature range between 21° and 28 °C and relative humidity less than 80% had significant association with VL in Kashi prefecture (Li and Zheng 2019); and also rainfall below 91.9 mm (Zheng et al. 2018). In the Mediterranean region of Thessaly, Greece, maximum temperature (32 ± 1 °C) accounted for 6% of the variance in an ecological niche model (ENM) for *Leishmania* infection (Giannakopoulos et al. 2016). In Africa, Ethiopia, a semiarid country, annual average temperatures between 20° and 37 °C (OR = 5.16) and annual rainfall below 766 mm were also predictors of

VL (Tsegaw et al. 2013). In Geradaf, Sudan, rainfall below 939 mm was the best predictor of VL incidence (Elnaiem et al. 2003b). In South America, Brazil, the incidence of VL increased in Tocantis with a mean night temperature range between 19° and 23 °C, humidity range between 8.42 and 16.72 g/kg, and annual rainfall between 1154.4 and 1710.2 mm (dos Reis et al. 2019). In São Paulo State, a rainfall range between 270 and 540 mm contributed the incidence of VL (da Paixão Sevá et al. 2017); other study in the same State found also positive association of VL cases with rainfall 1407 ± 123 mm but rainy days of 112 ± 18 were negative associated with VL cases (Oliveira et al. 2018),

Cutaneous leishmaniasis

Climate variables were also significant risk factors for CL studies; temperature was significant for 67% of 30 studies, rainfall for 40% of 30 studies, and relative humidity for 7 studies. In South America, a peak of incidence of CL was found in Chaparral, Colombia with a mean temperature of 20.6 ± 1.4 °C (Valderrama-Ardila et al. 2010), and a $\sim 16 \pm 5.7$ °C with all areas of incidence in the Andean region of Colombia (Pérez-Flórez et al. 2016). In addition, the same study in this Andean region found CL incidence was higher in areas where annual rainfall was 1841 ± 660.3 mm. In a geostatistical model of leishmaniasis incidence in Brazil, a 207–530 mm range of precipitation in the warmest quarter was an important risk factor for CL (Karagiannis-Voules et al. 2013).

In North Africa, a temperature of 9.4° to 22.1 °C contributed 20.7% of the variation in an Ecological Niche Model of the vector in Tunisia. Furthermore, in this country, the vector occurred in the driest quarter of the year with rainfall below to 37 mm (Chalghaf et al. 2016) and the relative humidity between 30 and 45% increase the CL incidence (Talmoudi et al. 2017). In addition, in Ethiopia, a temperature range between 17.2° and 23.8 °C was associated with CL occurrence (OR = 25.70) and annual rainfall between 903.4 and 1715.8 mm increased the risk of CL (OR = 2.67) (Seid et al. 2014).

In Isfahan, Iran, hot spot areas of CL presented annual temperature range between 15° and 19 °C, annual rainfall 5–20 mm, and relative humidity range between 27 and 36% (Ramezankhani et al. 2017b, 2018). In the same country, climate variables associated with the increase of CL incidence in Fars were rainy days 31 ± 13 and relative humidity of $47 \pm 17\%$ (Ali-Akbarpour et al. 2012); and in Khuzestan were temperatures of 34 ± 10 °C, rainy days from 30 to 65 days, and relative humidity of 10–78% (Azimi et al. 2017). However, in central region of Afghanistan, outdoor temperature of 2.16 °C was associated with increase of anthroponotic CL risk (Adegboye et al. 2019).

Discussion

Our analyses demonstrate that the incidence of leishmaniasis is influenced by a variety of environmental, landscape, and socioeconomic factors. In the first half of the twentieth century, the major risk factors for leishmaniasis were proximity to forest areas and distance from population centers (Southgate 1964; Moškovskij and Duhanina 1971; Ashford et al. 1973; Forattini et al. 1976). In the early 1990s, studies demonstrated that deforestation and development of rural settlements near forests increased VL and CL incidence (Montoya et al. 1990; Mott et al. 1990; Grimaldi and Tesh 1993), suggesting that the parasite life cycle can persist under altered ecological conditions by adopting peri-domestic sand flies as vectors and domestic animals as reservoirs. At present, people living in urban or peri-urban areas are at the greatest risk of infection (Machado-Coelho et al. 1999; Pigott et al. 2014a; Rocha et al. 2018). These populations have a high probability of contact with sand flies coupled with socioeconomic characteristics that make them particularly vulnerable to leishmaniasis.

We found a greater number of studies of VL relative to CL, consistent with previous reviews (Perilla-González et al. 2014). VL is lethal without treatment and is more common

in urban environments and as a result, has received more attention. However, CL was also present in periurban environments (Steffens 2010; Gomez-Barroso et al. 2015; de Oliveira et al. 2016) and without treatment could be associated with other health problems.

Below, we present a conceptual model to illustrate the interplay between risk factors analyzed in the studies included in this review and how such interactions can be used to understand the leishmaniasis transmission cycle (Fig. 5). The studies reviewed here not only identify several important factors related to leishmaniasis risk but also highlight how socioeconomic (Fig. 5a), landscape (Fig. 5b), and climatic factors (Fig. 5c) influence disease transmission.

Socioeconomic factors

In the Old World, Mediterranean, tropical, and arid regions have a long history of human intervention that has placed human populations close to the transmission foci. In contrast, environmental modifications in tropical areas of the New World are more recent and the percentage of the landscape that remains unmodified is still considerable in proportion to the modified area.

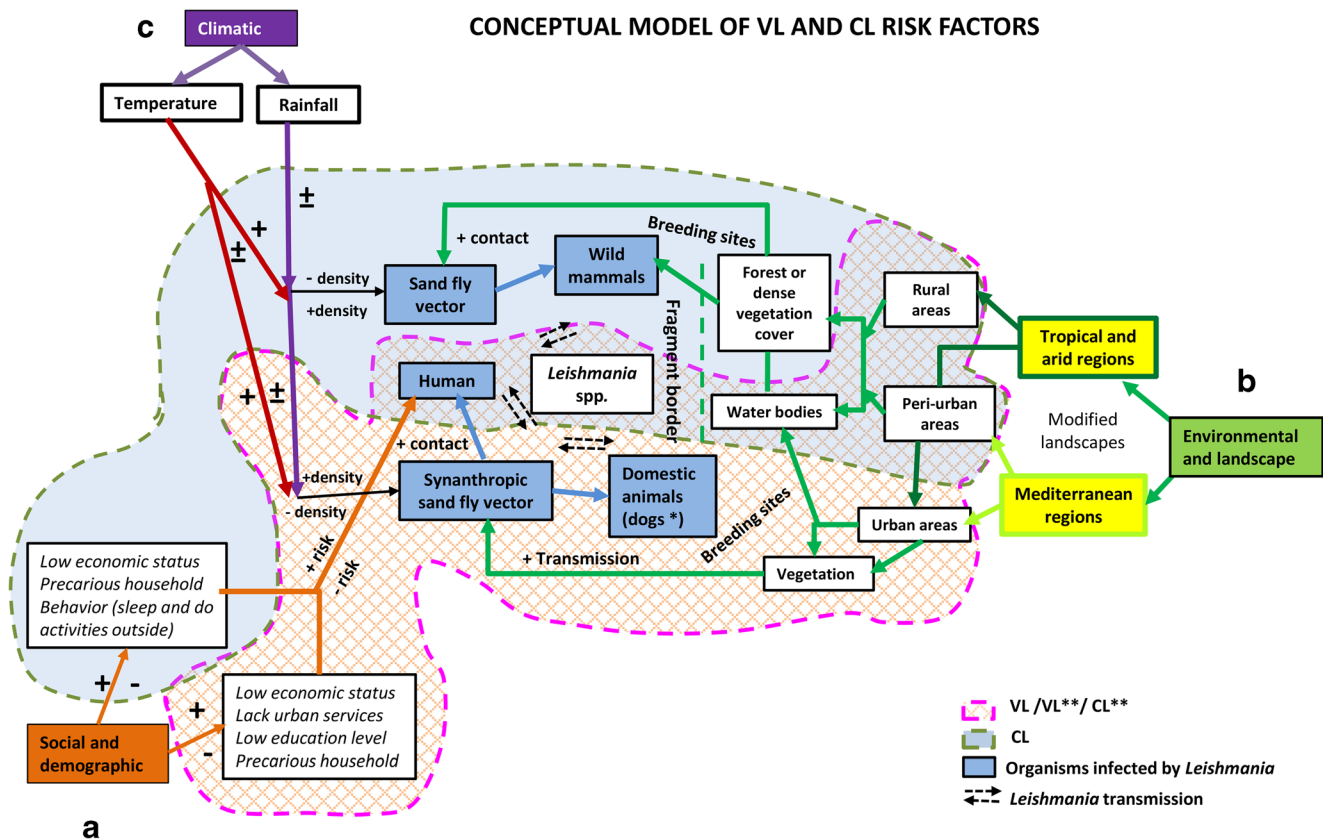


Fig. 5 Interaction of risk factors which influence the incidence of visceral (VL) and cutaneous leishmaniasis (CL). Arrows show factors that influence transmission: **a** orange = socioeconomic factors; **b** green = environmental factors; **c** climatic factors: brown = temperature; purple = rainfall.

(±) Represent intermediate values of temperature and rainfall. (*) Dogs are susceptible to some species responsible of VL especially in the New World. Segmented green line represents vegetation border between landscape components. (**) Anthroponotic leishmaniasis

Our analysis demonstrates that urbanization is linked with VL in the New World and with both clinical forms in the Old World. These observations are consistent with Pigott et al. (2014a), who found that urban land cover is associated with an increment of VL risk worldwide and for CL in the Old World. To become urbanized, leishmaniasis depends on the main vector species associated with transmission of the parasite. In the Old World where *Phlebotomus* spp. with synanthropic and anthrophilic behavior is the main vector, the transmission of VL and CL occurs in the peri-domestic environment in tropical rural areas while in Mediterranean regions, it occurs in urban areas and other peripheral urban settlements (Ready 2013). Similarly, in the tropical New World, *Lutzomyia longipalpis* is the main vector of *Leishmania infantum* responsible of VL. This vector species can survive in urban areas, even in the absence of surrounding forests (Lainson and Rangel 2005) and *L. infantum* has the dog as main reservoir, a species that is quite susceptible to infection. However, vector species of CL in the New World tropics usually remain in peripheral areas close to forest. In fact, the reservoirs for the parasites of CL are generally wild rodents that surround peri-urban areas (da Silva and Cunha 2007; Chavy et al. 2019).

Socioeconomic factors such as education level and poverty are frequently associated with malnutrition, poor housing, and lack of sanitary services (Navin et al. 1985; Adhikari et al. 2010b; Rijal et al. 2010; Maia et al. 2013, 2016; Abaker et al. 2017; Loiseau et al. 2019). These conditions can foster the occurrence of continued incidence of the disease (Ranjan et al. 2005; de Araújo and de Alencar Ximenes 2009; Adhikari et al. 2010a; Hasker et al. 2012; Bulstra et al. 2018). For example, precarious living conditions increase leishmaniasis risk because some types of household construction materials offer optimal conditions for sand fly development (Armijos et al. 1997; Reithinger et al. 2010; Singh et al. 2010; Ponte et al. 2011; Bamorovat et al. 2018). Phlebotomine adults find suitable resting and breeding sites in places where cracks or holes in the walls or damp floors are available (Bern et al. 2000; Ryan et al. 2006; Schenkel et al. 2006; Uranw et al. 2013). Lack of sanitation services can also attract wild or domestic reservoirs (Machado-Coelho et al. 1999; Arroub et al. 2012; Prestes-Carneiro et al. 2019) or be a potential breeding site for sand flies (Costa et al. 2005; Moreno et al. 2005).

All of these factors could increase the risk of leishmaniasis and also allow the development of the disease in infected individuals, especially if medical services are inadequate (Alvar et al. 2006; Perilla-González et al. 2014; Pigott et al. 2014a; Rodrigues et al. 2019) and households lack water supply (de Almeida et al. 2011; Lopez et al. 2018; Godana et al. 2019), increasing the prevalence in areas with these characteristics (Boelaert et al. 2009). Moreover, in areas where leishmaniasis overlaps with AIDS or other diseases, the risk of

leishmaniasis infection increases considerably (Desjeux 2001; Melchior et al. 2017; Lima et al. 2018). Such overlap is a risk factor in urban areas of the Mediterranean region of Europe where the re-emergence of VL and CL has been caused by an increase in the number of immunosuppressed people rather than lack of sanitation and health services (Steffens 2010).

A high number of individuals per dwelling could also reflect low socioeconomic status (Shah 2005; Alves et al. 2016). Rising population densities in peripheral neighborhoods could attract more sand flies because human CO₂ emissions attract the flies (Chaniotis 1983; Campbell-Lendrum et al. 1999; Chavy et al. 2019). Migration from rural to urban areas (Alcáis et al. 1997; Monteiro et al. 2009; Lemma et al. 2015; Abdulla et al. 2018) and the exponential population growth in peri-urban areas have increased the number of dwellings lacking sanitation services (Madalosso et al. 2012; Toledo et al. 2017; Hernández et al. 2019).

In addition, characteristics of population as age and gender were also associated as leishmaniasis risk factors. Adult males were the most exposed at sand flies bites due their occupational activities (Abaker et al. 2017; Nazari et al. 2017; Eid et al. 2018). Work in agriculture, hunting, or timber collection inside the forest especially at the end of the afternoon exposes individuals to sand fly bites, increasing the incidence of the disease (Jones et al. 1987; Espejo et al. 1989; Weigle et al. 1993; Azene et al. 2017; Bellali et al. 2017; Guerra et al. 2019). Sand flies are active at twilight and night; less so at dawn when the decrease of temperature drives them to their natural hiding places (Forattini et al. 1976). Sleeping outdoors or without protection against sand flies bites increments disease risk (Davies et al. 1997; Barnett et al. 2005; Bashaye et al. 2009; Nackers et al. 2015; Kumar et al. 2017). These conditions are generally more common in rural areas or areas close to forest (Almeida and Werneck 2014; Iddawela et al. 2018).

Likewise, disease risk increase with the presence of mammals that can act as potential reservoirs of the parasite. Wild, synanthropic, or domestic animals could be part of the cycle (Beier et al. 1986; Oliveira et al. 2006; Khanal et al. 2010; Cardoso et al. 2015; De Araujo et al. 2016). The presence of dogs in households is considered a high risk factor for the incidence of VL especially in the New World (Votýpka et al. 2012). Dogs are very susceptible to infection, develop the disease, and act as a source of further infection of *Leishmania infantum* one of the main agents responsible of VL in the New World, and both VL and CL in the Old World (Faris et al. 1988; Dantas-Torres et al. 2012; Bsrat et al. 2015, 2018). Although cattle can increase the risk (Ullah et al. 2016; Bhowmick and Khanum 2017; Ghatee et al. 2018), they may also decrease leishmaniasis (Bern et al. 2010) by serving as the principal source of sand fly blood meal and diverting bites to humans (Bern et al. 2005; Kolaczinski et al. 2008).

More research to establish control measures is needed to understand the influence of poverty on leishmaniasis occurrence (Bern et al. 2010; Rodríguez-Morales et al. 2010; El Alem et al. 2018). In the absence of control measures, other factors such as high educational level, good nutrition, and high wealth level cannot by themselves reduce the risk of infection. Furthermore, without the presence of the vector at high densities, new settlements in periurban areas are not always at high risk for leishmaniasis (Alves et al. 2016; Gijón-Robles et al. 2018).

Environmental and landscape factors

Conversion of natural forest to other land uses in the last decades has led to habitat fragmentation and altered landscape composition (Wade et al. 2003). The spread of the vector and disease at macro scales is associated with migration and expansion of human population into natural areas, creation of roadways, energy networks, new farm lands, and poorly planned urban development (Cardim et al. 2013; Afonso et al. 2017; Gutierrez et al. 2017; Oliveira et al. 2018). These changes in the landscape increase contact of human populations with the edges of vegetation areas which shelter sand fly vectors (Patz et al. 2004). New settlements near forests act as foci of leishmaniasis enabling the domestication of the cycle. At the same time, expansion of agricultural crops provides a new food source for natural reservoirs of leishmaniasis such as rodents (Zorrilla et al. 2005; Dawit et al. 2013; Ranasinghe et al. 2013; Carrada Figueroa Gdel et al. 2014; de Oliveira et al. 2016; Gebremichael Tedla et al. 2018).

In human-modified landscapes, fragments of vegetation close to dwellings play an important role in the transmission cycle. Vegetation areas provide the environmental conditions for sand fly breeding sites and development while at the same time act as shelters of wild reservoirs (Negera et al. 2008; Ocampo et al. 2012; de Almeida et al. 2014; de Santana Martins 2015; Temponi et al. 2018; Melo et al. 2018). Populations of many phlebotomine species increase from secondary forest to mature forests (Rutledge and Gupta 2002; Gil et al. 2010; Kariyawasam et al. 2015) except for the species that have adapted to urban environments and do not need dense vegetation to survive such as *Lutzomyia longipalpis* (Salomón et al. 2015) and *Phlebotomus argentipes* (Bhunja et al. 2010a, 2012).

High values of NDVI or vegetation near incidence areas found in several studies suggest proximity to natural habitats connects human dwellings with the vector breeding sites (Aparicio and Bitencourt 2004; Cerbino Neto et al. 2009; Gomez-Barroso et al. 2015; Menezes et al. 2016; Artun and Kavur 2017). The presence of vegetation or forest areas near houses increase the number of sand flies in peri-domestic areas and the probability of being infected (Miranda et al.

1998; Andrade-Narvaez et al. 2003; Dias et al. 2007). Indeed, studies that analyzed the abundance of the vector at different distances from the forest to households of infected people found the number of cases decreased with distance (Werneck et al. 2002; Feliciangeli et al. 2006). The flight range of sand flies is around 200 m, with *P. argentipes* and *P. orientalis* having flight ranges of ca. 500 m and those of *L. longipalpis* and *P. caucasicus* 1000 m or more (Rutledge and Gupta 2002). However, vegetation is a risk factor in tropical forest areas and Mediterranean forest, but not in arid areas where vegetation is not dense and the burrows of mammals, caves, crevice, termite hills, and walls with cracks may serve as breeding sites of sand flies (Holakouie-Naieni et al. 2017; Lotukoi 2017; Ramezankhani et al. 2017a; Alebie et al. 2019; Dulacha et al. 2019).

The presence of waterbodies is also related to vector abundance and its distribution (Yadon et al. 2003; Schenkel et al. 2006; Nilforoushzadeh et al. 2014; de Oliveira et al. 2016; Rajabi et al. 2016b), possibly because waterbodies provide the air moisture necessary for sand fly breeding (Bhunja et al. 2011). The effects of soil type on disease incidence may also reflect moisture conditions. Studies showed that soil types characterized by water retention as fluvisols, the typical soil of river areas, soils with some organic material as inceptisol, planosol which maintain moisture, torrifluvents, humid haplocalcid soils with torriorthents, and clayey soils which also retain water were associated with high disease incidence, possibly by facilitating larval development of sand flies (El-naïem et al. 2003a; Sharma and Singh 2008).

Because proximity to forests and waterbodies have been associated with disease incidence, control measures in recent years have focused on forest clearing and wetland drainage near incidence areas (Wood et al. 2014). However, we do not advocate that deforestation and wetland draining is the solution for decreasing the risk of leishmaniasis. More studies are necessary to clarify how exposure to the disease agent would change the structure, composition, or function of landscape changes (Myers et al. 2013), seeking environmentally friendly alternatives of disease control.

In our review, we could not identify a range of altitudes significant for leishmaniasis incidence. Clearly, altitude is related with other environmental features that influence vector distribution (Quintana et al. 2012; Ferro et al. 2015; Prudhomme et al. 2015; Mokhtari et al. 2016; Moradi-Asl et al. 2017; Al-Warid et al. 2019). Likewise, topographic characteristics such as slope show a significant impact on the presence and abundance of vectors. However, it is unclear whether these effects simply reflect the indirect effects of temperature shifts with elevation or soil moisture with topography highlighting the need for more studies that can clarify the importance of these factors in the genesis and transmission of the disease.

Climatic factors

Climatic conditions are generally important risk factors for vector-borne diseases (Cardenas et al. 2006). Their effects on leishmaniasis vary according to geographic area and depend on the clinical form and vector species. Studies in Mediterranean, tropical, and arid regions suggest that sand flies thrive between 19° and 30 °C (Kassem et al. 2012; de Souza et al. 2015; Giannakopoulos et al. 2016; Pérez-Flórez et al. 2016; Amro et al. 2017; Abdullah et al. 2017; Artun and Kavur 2018; Moradiasl et al. 2018; Galgamuwa et al. 2018) with a relative humidity above 30% (Gao and Cao 2019), and bite at temperatures between 20° and 30 °C (Rutledge and Gupta 2002). Temperatures over 30 °C and without enough relative humidity negatively affect sand fly population density (Karagiannis-Voules et al. 2013; Moo Llanes 2016). However, in central Afghanistan, a cold outdoor temperature range between 2° and 6 °C increases the incidence of anthroponotic CL, because *Phlebotomus sergenti* the main vector of *L. tropica* is anthropophilic and individuals are mostly indoors where there is a temperature range between 21° and 26 °C (Adegboye et al. 2019).

Rainfall was also associated with leishmaniasis transmission and vector abundance. High rainfall and relative humidity (Elnaiem et al. 2003b; Pérez-Flórez et al. 2016) increase primary productivity (Salomón et al. 2004; Ben-Ahmed et al. 2009; Souza et al. 2012) of forest vegetation which provides food and burrows for reservoirs (Chalghaf et al. 2016), providing an ideal environment for sand flies (Andrade-Narvaez et al. 2003; Quintana et al. 2012; Adegboye and Adegboye 2017). The vectors of *Leishmania* involved in each of the two clinical forms have a different rainfall range around the world. Based on the literature reviewed, we cannot specify a range of precipitation as we did with temperature. However, some modeling analysis report a high precipitation index is an important environmental factor for cutaneous leishmaniasis incidence in the tropics (Chaves and Pascual 2006; Ali-Akbarpour et al. 2012) because the vectors related to this clinical form are associated with the presence of dense vegetation. However, according to some studies, high precipitation is not favorable for visceral leishmaniasis in the tropics (Thompson et al. 2002; Viana et al. 2011; Karagiannis-Voules et al. 2013; Pigott et al. 2014b), but other studies positively correlated rainfall indices with VL (da Paixão Sevá et al. 2017; Oliveira et al. 2018; dos Reis et al. 2019). Taking into account that VL in tropical regions occurs in highly urbanized areas, we can hypothesize that moderate levels of precipitation can foster incidence areas by maintaining humidity in the environment, and sand fly vectors are able to thrive in climatic variations in this regions. Nevertheless, without the protection of vegetation, heavy precipitation could decimate sand fly populations.

There are some limitations for this systematic review. In order to do a comprehensive descriptive review, we did not restrict our inclusion criteria according to the study design (e.g., cohort studies, case-control studies, spatial analysis, ecological niche modelling), and all the studies which analyze risk factors of VL and CL were included. Some studies did not describe the range values of environmental variables significant. Some studies in regions where anthroponotic and zoonotic leishmaniasis coexists did not specify the type of transmission, so we considered only the clinical form. However, the summary of information assessed provides a data-based assessment of the degree to which recent changes in land use, the size and distribution of human populations, and a changing climate have influenced the occurrence of transmission dynamics for the disease.

Conclusion

Our review describes the complexity of transmission and incidence of a disease that presents two main clinical forms and can exist at a broad range of environmental and climatic conditions. We also highlight how complex interaction between risk factors can exacerbate or moderate the incidence of leishmaniasis.

High disease incidence is associated with several environmental, climatic, and socioeconomic conditions. Transmission patterns are similar in all regions, requiring human contact with vegetation areas that harbor reservoir vectors and mammals under warm climatic conditions. Differences among clinical forms and regions depend on the species of vectors involved in each type of leishmaniasis and if these are able to easily adapt to urban environments (e.g., VL in the tropics, arid and Mediterranean regions) or depend on less disturbed environments (e.g., CL in the Neotropics). More research that analyze the interactions of risks factors and how they vary across vector, reservoir species, and environmental conditions in countries where the disease is endemic, is needed. Thus, developing effective control measures will be possible and also require a better understanding of the likely impacts of future climatic conditions on the transmission cycle.

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Compliance with Ethical Standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Abaker AS, Mohammed AA, Elawad AE (2017) Socioeconomic and behavioral risk factors for infection of visceral leishmaniasis Gedaref State - Sudan. *Glob J Med Public Health* 6:1–7
- Abdulla QB, Shabila NP, Al-Hadithi TS (2018) An outbreak of cutaneous leishmaniasis in erbil governorate of Iraqi Kurdistan region in 2015. *J Infect Dev Ctries* 12:600–607. <https://doi.org/10.3855/JIDC.10306>
- Abdullah AYM, Dewan A, Shogib MRI et al (2017) Environmental factors associated with the distribution of visceral leishmaniasis in endemic areas of Bangladesh: modeling the ecological niche. *Trop Med Health* 45:13. <https://doi.org/10.1186/s41182-017-0054-9>
- Adegboye OA, Adegboye M (2017) Spatially correlated time series and ecological niche analysis of cutaneous leishmaniasis in Afghanistan. *Int J Environ Res Public Health* 14:1–14. <https://doi.org/10.3390/ijerph14030309>
- Adegboye MA, Olumoh J, Saffary T et al (2019) Effects of time-lagged meteorological variables on attributable risk of leishmaniasis in central region of Afghanistan. *Sci Total Environ* 685:533–541. <https://doi.org/10.1016/j.scitotenv.2019.05.401>
- Adhikari SR, Supakankunti S, Khan MM (2010a) Kala azar in Nepal: Estimating the effects of socioeconomic factors on disease incidence. *Kathmandu Univ Med J* 8(1):73–79
- Adhikari SR, Supakankunti S, Khan MM (2010b) Incidence of kala-azar in Nepal: estimating the effects of individual and household characteristics. *Trans R Soc Trop Med Hyg* 104:720–725. <https://doi.org/10.1016/j.trstmh.2010.08.013>
- Akhoundi M, Kuhls K, Cannet A et al (2016) A historical overview of the classification, evolution, and dispersion of Leishmania parasites and sandflies. *PLoS Negl Trop Dis* 10:1–40. <https://doi.org/10.1371/journal.pntd.0004349>
- Alcáiz A, Abel L, David C et al (1997) Risk factors for onset of cutaneous and mucocutaneous leishmaniasis in Bolivia. *Am J Trop Med Hyg* 57:79–84
- Alebie G, Worku A, Yohannes S et al (2019) Epidemiology of visceral leishmaniasis in Shebelle Zone of Somali Region, eastern Ethiopia. *Parasit Vectors* 12:1–10. <https://doi.org/10.1186/s13071-019-3452-5>
- Alemayehu B, Alemayehu M (2017) Leishmaniasis: a review on parasite, vector and reservoir host. *Health Sci J* 11:1–6. <https://doi.org/10.21767/1791-809X.1000519>
- Ali-Akbarpour M, Mohammadbeigi A, Tabatabaee SHR, Hatam G (2012) Spatial analysis of eco-environmental risk factors of cutaneous leishmaniasis in Southern Iran. *J Cutan Aesthet Surg* 5:30. <https://doi.org/10.4103/0974-2077.94338>
- Almeida AS, Werneck GL (2014) Prediction of high-risk areas for visceral leishmaniasis using socioeconomic indicators and remote sensing data. *Int J Health Geogr* 13:7. <https://doi.org/10.1186/1476-072X-13-13>
- Alvar J, Yactayo S, Bern C (2006) Leishmaniasis and poverty. *Trends Parasitol* 22:552–557. <https://doi.org/10.1016/j.pt.2006.09.004>
- Alvar J, Vélez ID, Bern C et al (2012) Leishmaniasis worldwide and global estimates of its incidence. *PLoS One* 7:1–12. <https://doi.org/10.1371/journal.pone.0035671>
- Alves EB, Costa CHN, de Carvalho FAA et al (2016) Risk profiles for Leishmania infantum infection in Brazil. *Am J Trop Med Hyg* 94:1276–1281. <https://doi.org/10.4269/ajtmh.15-0513>
- Al-Warid HS, Al-Saqur IM, Kadhem AJ et al (2019) Spatial and demographic aspects of kala-azar (Visceral leishmaniasis) in Iraq during 2011–2013. *Trop Biomed* 36:22–34
- Amro A, Al-Dwibe H, Gashout A et al (2017) Spatiotemporal and molecular epidemiology of cutaneous leishmaniasis in Libya. *PLoS Negl Trop Dis* 11:e0005873. <https://doi.org/10.1371/journal.pntd.0005873>
- Andrade-Narvaez FJ, Canto Lara SB, Van Wylsberghe NR et al (2003) Seasonal transmission of Leishmania (Leishmania) mexicana in the State of Campeche, Yucatan Peninsula, Mexico. *Mem Inst Oswaldo Cruz* 98:995–998. <https://doi.org/10.1590/S0074-02762003000800002>
- Antonioli SAC, Torres TG, Filho ACP, Tolezano JE (2007) Spatial analysis of American visceral leishmaniasis in Mato Grosso do Sul State, Central Brazil. *J Inf Secur* 54:509–514. <https://doi.org/10.1016/j.jinf.2006.08.004>
- Aparicio C, Bitencourt MD (2004) Modelagem espacial de zonas de risco da leishmaniose tegumentar americana. *Rev Saude Publica* 38:511–516. <https://doi.org/10.1590/S0034-89102004000400005>
- Argaw D, Mulugeta A, Herrero M et al (2013) Risk factors for visceral Leishmaniasis among residents and migrants in Kafta-Humera, Ethiopia. *PLoS Negl Trop Dis* 7:9. <https://doi.org/10.1371/journal.pntd.0002543>
- Armijos RX, Weigel MM, Izurieta R et al (1997) The epidemiology of cutaneous leishmaniasis in subtropical Ecuador. *Tropical Med Int Health* 2:140–152. <https://doi.org/10.1046/j.1365-3156.1997.d01-236.x>
- Arroub H, Alaoui A, Lemrani M, Habbari K (2012) Cutaneous Leishmaniasis in Fom Jamâa (Azilal, Morocco): micro-environmental and socio-economical risk factors. *J Agric Soc Sci* 8:1813–2235
- Artun O, Kavur H (2017) Geographical information systems in determination of spatial factors in cutaneous leishmaniasis cases distribution, in Adana, Turkey. *Sci Pap E-I Reclam Earth Obs Surv Environ Eng* 6:141–146
- Artun O, Kavur H (2018) Prediction of cutaneous Leishmaniasis epidemiology in mersin using ecological niche modeling. *Turkiye parazitolojii Derg* 42:191–195. <https://doi.org/10.5152/tpd.2018.5924>
- Ashford RW (1996) Leishmaniasis reservoirs and their significance in control. *Clin Dermatol* 14:523–532. [https://doi.org/10.1016/0738-081X\(96\)00041-7](https://doi.org/10.1016/0738-081X(96)00041-7)
- Ashford RW, Bray MA, Hutchinson MP, Bray RS (1973) The Epidemiology of cutaneous leishmaniasis in Ethiopia. *Trans R Soc Trop Med Hyg* 67:568–601
- Assefa A (2018) Leishmaniasis in Ethiopia: a systematic review and meta-analysis of prevalence in animals and humans. *Heliyon* 4:e00723. <https://doi.org/10.1016/j.heliyon.2018.e00723>
- Azene W, Sissay M, Kebede A, Gashaw F (2017) Visceral leishmaniasis and associated risk factors in Libo Kemkem, Bura Egzi Abhier Ab Kebele, Northwestern Ethiopia. *EC Microbiol* 7:162–172
- Azimi F, Shirian S, Jangjoo S et al (2017) Impact of climate variability on the occurrence of cutaneous leishmaniasis in Khuzestan Province, southwestern Iran. *Geospat Health* 12:15–22. <https://doi.org/10.4081/gh.2017.478>
- Badaró R (1988) Progress of research in visceral leishmaniasis in the endemic area of Jacobina—Bahia 1934–1989. *Rev Soc Bras Med Trop* 21:159–164
- Bamorovat M, Sharifi I, Aflatoonian MR et al (2018) Risk factors for anthroponotic cutaneous leishmaniasis in unresponsive and responsive patients in a major focus, southeast of Iran. *PLoS One* 13:e0192236. <https://doi.org/10.1371/journal.pone.0192236>
- Barnett P, Singh SP, Bern C et al (2005) Virgin soil: the spread of visceral leishmaniasis into Uttar Pradesh, India. *Am J Trop Med Hyg* 73:720–725

- Bashaye S, Nombela N, Argaw D et al (2009) Risk factors for visceral leishmaniasis in a new epidemic site in Amhara region, Ethiopia. *Am J Trop Med Hyg* 81:34–39
- Bates PA, Depaquit J, Galati EAB et al (2015) Recent advances in phlebotomine sand fly research related to leishmaniasis control. *Parasit Vectors* 8:1–8. <https://doi.org/10.1186/s13071-015-0712-x>
- Bavia ME, Carneiro DDMT, Gurgel Hda C et al (2005) Remote sensing and geographic information systems and risk of American visceral leishmaniasis in Bahia, Brazil. *Parasitologia* 47:165–169
- Beier JC, El Sawaf BM, Merdan AI et al (1986) Sand flies (Diptera : Psychodidae) associated with visceral leishmaniasis in El Agamy, Alexandria Governorate, Egypt. I. Population ecology. *J Med Entomol* 23:600–608
- Bellali H, Chemak F, Nouri I et al (2017) Zoonotic cutaneous leishmaniasis prevalence among farmers in Central Tunisia, 2014. *J Agromed* 22:244–250. <https://doi.org/10.1080/1059924X.2017.1318725>
- Belo VS, Werneck GL, Barbosa DS et al (2013) Factors associated with visceral leishmaniasis in the Americas: a systematic review and meta-analysis. *PLoS Negl Trop Dis* 7:12. <https://doi.org/10.1371/journal.pntd.0002182>
- Ben-Ahmed K, Aoun K, Jeddi F et al (2009) Visceral leishmaniasis in Tunisia: spatial distribution and association with climatic factors. *Am J Trop Med Hyg* 81:40–45
- Bern C, Joshi AB, Jha SN et al (2000) Factors associated with visceral leishmaniasis in Nepal: bed-net use is strongly protective. *Am J Trop Med Hyg* 63:184–188
- Bern C, Hightower AW, Chowdhury R et al (2005) Risk factors for kala-azar in Bangladesh. *Emerg Infect Dis* 11:655–662. <https://doi.org/10.3201/eid1105.040718>
- Bern C, Courtenay O, Alvar J (2010) Of cattle, sand flies and men: a systematic review of risk factor analyses for South Asian visceral leishmaniasis and implications for elimination. *PLoS Negl Trop Dis* 4:1–10. <https://doi.org/10.1371/journal.pntd.0000599>
- Bhowmick AR, Khanum H (2017) Prevalence of visceral leishmaniasis, risk factors and associated disorders: knowledge of inhabitants and professionals in Fulbaria, Mymensingh. *Bangladesh J Zool* 45:73–83. <https://doi.org/10.3329/bjz.v45i1.34197>
- Bhunias GS, Kesari S, Jeyaram A et al (2010a) Influence of topography on the endemicity of Kala-azar: a study based on remote sensing and geographical information system. *Geospat Health* 4:155–165. <https://doi.org/10.4081/gh.2010.197>
- Bhunias GS, Kumar V, Kumar AJ et al (2010b) The use of remote sensing in the identification of the eco-environmental factors associated with the risk of human visceral leishmaniasis (kala-azar) on the Gangetic plain, in north-eastern India. *Ann Trop Med Parasitol* 104:35–53. <https://doi.org/10.1179/136485910X12607012373678>
- Bhunias GS, Kesari S, Chatterjee N et al (2011) Incidence of visceral leishmaniasis in the Vaishali district of Bihar, India: spatial patterns and role of inland water bodies. *Geospat Health* 5:205–215. <https://doi.org/10.4081/gh.2011.173>
- Bhunias GS, Chatterjee N, Kumar V et al (2012) Delimitation of kala-azar risk areas in the district of Vaishali in Bihar (India) using a geo-environmental approach. *Mem Inst Oswaldo Cruz* 107:609–620. <https://doi.org/10.1590/S0074-02762012000500007>
- Boelaert M, Meheus F, Sanchez A et al (2009) The poorest of the poor: a poverty appraisal of households affected by visceral leishmaniasis in Bihar, India. *Tropical Med Int Health* 14:639–644. <https://doi.org/10.1111/j.1365-3156.2009.02279.x>
- Borges BKA, Silva JA, JPA H et al (2009) Presença de animais associada ao risco de transmissão da leishmaniose visceral em humanos em Belo Horizonte, Minas Gerais. *Arq Bras Med Vet e Zootec* 61: 1035–1043. <https://doi.org/10.1590/S0102-09352009000500004>
- Bsrat A, Berhe M, Balkew M et al (2015) Epidemiological study of cutaneous leishmaniasis in Saesie Tsaeda-embra district, eastern Tigray, Northern Ethiopia. *Parasit Vectors* 8:1–9. <https://doi.org/10.1186/s13071-015-0758-9>
- Bsrat A, Berhe M, Gadissa E et al (2018) Serological investigation of visceral Leishmania infection in human and its associated risk factors in Welkait District, Western Tigray, Ethiopia. *Parasite Epidemiol Control* 3:13–20. <https://doi.org/10.1016/j.parepi.2017.10.004>
- Bulstra CA, Le Rutte EA, Malaviya P et al (2018) Visceral leishmaniasis: spatiotemporal heterogeneity and drivers underlying the hotspots in Muzaffarpur, Bihar, India. *PLoS Negl Trop Dis* 12:1–21. <https://doi.org/10.1371/journal.pntd.0006888>
- Campbell-Lendrum DH, Pinto MC, Brandão-Filho SP et al (1999) Experimental comparison of anthropophily between geographically dispersed populations of *Lutzomyia whitmani* (Diptera: Psychodidae). *Med Vet Entomol* 13:299–309. <https://doi.org/10.1046/j.1365-2915.1999.00174.x>
- Cardenas R, Sandoval CM, Rodriguez-Morales AJ, Franco-Paredes C (2006) Impact of climate variability in the occurrence of leishmaniasis in Northeastern Colombia. *Am J Trop Med Hyg* 75:273–277
- Cardim MFM, Rodas LAC, Dibo MR et al (2013) Introduction and expansion of human American visceral leishmaniasis in the state of Sao Paulo, Brazil, 1999–2011. *Rev Saude Publica* 47:1–9. <https://doi.org/10.1590/S0034-8910.2013047004454>
- Cardim MFM, Guirado MM, Dibo MR, Neto FC (2016) Visceral leishmaniasis in the state of Sao Paulo, Brazil: spatial and space-time analysis. *Rev Saude Publica* 50:1–11. <https://doi.org/10.1590/S1518-8787.2016050005965>
- Cardoso RM, de Araújo NNSL, Romero GAS et al (2015) Expanding the knowledge about Leishmania species in wild mammals and dogs in the Brazilian savannah. *Parasit Vectors* 8:1–8. <https://doi.org/10.1186/s13071-015-0780-y>
- Carrada Figueroa Gdel C, Leal Ascencio VJ, Jiménez Sastré A, López Álvarez J (2014) Transmission of cutaneous leishmaniasis associated with cacao (*Theobroma cacao*) plantations in Tabasco. *Gac Med Mex* 150:499–508
- Cerbino Neto J, Werneck GL, Costa CHN (2009) Factors associated with the incidence of urban visceral leishmaniasis: an ecological study in Teresina, Piauí State, Brazil. *Cad Saude Publica* 25:1543–1551. <https://doi.org/10.1590/S0102-311X2009000700012>
- Chalhaf B, Chlif S, Mayala B et al (2016) Ecological niche modeling for the prediction of the geographic distribution of cutaneous leishmaniasis in Tunisia. *Am J Trop Med Hyg* 94:844–851. <https://doi.org/10.4269/ajtmh.15-0345>
- Chaniotis BN (1983) Improved trapping of phlebotomine sand flies (Diptera: Psychodidae) in light traps supplemented with dry ice in a neotropical rain forest. *J Med Entomol* 20:222–223
- Chaves LF, Pascual M (2006) Climate cycles and forecasts of cutaneous leishmaniasis, a nonstationary vector-borne disease. *PLoS Med* 3: 1320–1328. <https://doi.org/10.1371/journal.pmed.0030295>
- Chavy A, Ferreira Dales Nava A, Luz SLB et al (2019) Ecological niche modelling for predicting the risk of cutaneous leishmaniasis in the Neotropical moist forest biome. *PLoS Negl Trop Dis* 13:e0007629. <https://doi.org/10.1371/journal.pntd.0007629>
- Costa CH, Werneck GL, Rodrigues L Jr et al (2005) Household structure and urban services: neglected targets in the control of visceral leishmaniasis. 99:229–236. <https://doi.org/10.1179/136485905X28018>
- da Paixão Sevã A, Mao L, Galvis-Ovallos F et al (2017) Risk analysis and prediction of visceral leishmaniasis dispersion in São Paulo State, Brazil. *PLoS Negl Trop Dis* 11:1–17. <https://doi.org/10.1371/journal.pntd.0005353>
- da Silva LMR, Cunha PR (2007) A urbanização da leishmaniose tegumentar americana no município de Campinas – São Paulo (SP) e região: Magnitude Campinas – São Paulo (SP) and region: problems and challenges. *An Bras Dermatol* 82:515–519. <https://doi.org/10.1590/S0365-05962007000600003>
- Dantas-Torres F, Brandão-Filho SP (2006) Visceral leishmaniasis in Brazil: revisiting paradigms of epidemiology and control. *Rev Inst*

- Med Trop Sao Paulo 48:151–156. <https://doi.org/10.1590/S0036-46652006000300007>
- Dantas-Torres F, Solano-Gallego L, Baneth G et al (2012) Canine leishmaniosis in the Old and New Worlds: unveiled similarities and differences. *Trends Parasitol* 28:531–538. <https://doi.org/10.1016/j.pt.2012.08.007>
- Davies CR, Llanos-Cuentas EA, Campos P et al (1997) Cutaneous leishmaniasis in the peruvian Andes: risk factors identified from a village cohort study. *Am J Trop Med Hyg* 56:85–95
- Dawit G, Girma Z, Simenew K (2013) A review on biology, epidemiology and public health significance of leishmaniasis. *J Bacteriol Parasitol* 4:7. <https://doi.org/10.4172/2155-9597.1000166>
- de Almeida AS, Medronho Rde A, Werneck GL (2011) Identification of risk areas for visceral leishmaniasis in Teresina, Piauí State, Brazil. *Am J Trop Med Hyg* 84:681–687. <https://doi.org/10.4269/ajtmh.2011.10-0325>
- de Almeida AS, Werneck GL, da Costa Resendes AP (2014) Classificação orientada a objeto de imagens de sensoriamento remoto em estudos epidemiológicos sobre leishmaniose visceral em área urbana. *Cad Saude Publica* 30:1639–1653. <https://doi.org/10.1590/0102-311X00059414>
- de Araújo PF, de Alencar Ximenes RA (2009) Sociodemographic and environmental risk factors for American cutaneous leishmaniasis (ACL) in the State of Alagoas, Brazil. *Am J Trop Med Hyg* 81:195–201
- de Araújo VEM, Pinheiro LC, de Mattos Almeida MC et al (2013) Relative risk of visceral leishmaniasis in Brazil: a spatial analysis in urban area. *PLoS Negl Trop Dis* 7:9. <https://doi.org/10.1371/journal.pntd.0002540>
- De Araujo AR, Portela NC, Feitosa APS et al (2016) Risk factors associated with american cutaneous leishmaniasis in an endemic area of Brazil. *Rev Inst Med Trop Sao Paulo* 58:2–7. <https://doi.org/10.1590/S1678-9946201658086>
- de Freitas Rocha AT, de Espindola GM, Soares MRA et al (2018) Visceral leishmaniasis and vulnerability conditions in an endemic urban area of Northeastern Brazil. *Trans R Soc Trop Med Hyg* 112:317–325. <https://doi.org/10.1093/trstmh/try058>
- de Oliveira EF, Silva EA, Fernandes CE et al (2012) Biotic factors and occurrence of *Lutzomyia longipalpis* in endemic area of visceral leishmaniasis, Mato Grosso do Sul, Brazil. *Mem Inst Oswaldo Cruz* 107:396–401. <https://doi.org/10.1590/S0074-02762012000300015>
- de Oliveira IP, da Fonseca AH, Silveira AK et al (2016) Spatial analysis of areas likely to harbor American cutaneous leishmaniasis in Seropédica, Rio de Janeiro State, Brazil. *Semin Ciências Agrárias* 37:193–202. <https://doi.org/10.5433/1679-0359.2016v37n1p193>
- de Santana Martins M (2015) The use of geographic information systems and ecological niche modeling to map transmission risk for visceral leishmaniasis in Bahia. Louisiana State University, Brazil
- de Souza RAF, Andreoli RV, Kayano MT, Carvalho AL (2015) American cutaneous leishmaniasis cases in the metropolitan region of Manaus, Brazil: association with climate variables over time. *Geospat Health* 10:40–47. <https://doi.org/10.4081/gh.2015.314>
- Desjeux P (2001) The increase in risk factors for leishmaniasis worldwide. *Trans R Soc Trop Med Hyg* 95:239–243. [https://doi.org/10.1016/S0035-9203\(01\)90223-8](https://doi.org/10.1016/S0035-9203(01)90223-8)
- Desjeux P (2004) Leishmaniasis: Current situation and new perspectives. *Comp Immunol Microbiol Infect Dis* 27:305–318. <https://doi.org/10.1016/j.cimid.2004.03.004>
- Dias ES, França-Silva JC, Costa J et al (2007) Flebotomíneos (Diptera : Psychodidae) de um foco de leishmaniose tegumentar no Estado de Minas Gerais Sandflies (Diptera : Psychodidae) in an outbreak of cutaneous leishmaniasis in the State of Minas Gerais. *Rev Soc Bras Med Trop* 40:49–52
- dos Reis LL, da Silva Balieiro AA, Fonseca FR, Gonçalves MJF (2019) Leishmaniose visceral e sua relação com fatores climáticos e ambientais no Estado do Tocantins, Brasil, 2007 a 2014. *Cad Saude Publica* 35:e00047018. <https://doi.org/10.1590/0102-311x00047018>
- dos Santos Afonso MM, de Miranda Chaves SA, de Avelar Figueiredo Mafra Magalhães M, et al (2017) Ecoepidemiology of American visceral Leishmaniasis in Tocantins State, Brazil: factors associated with the occurrence and spreading of the vector *Lutzomyia (Lutzomyia) longipalpis* (Lutz & Neiva, 1912) (Diptera: Psychodidae: Phlebotominae). In: *The Epidemiology and Ecology of Leishmaniasis*. InTech, p 91
- Dujardin J-C (2006) Risk factors in the spread of leishmaniasis: towards integrated monitoring? *Trends Parasitol* 22:4–6. <https://doi.org/10.1016/j.pt.2005.11.004>
- Dulacha D, Mwatha S, Lomurukai P et al (2019) Epidemiological characteristics and factors associated with Visceral Leishmaniasis in Marsabit County, Northern Kenya. *J Interv Epidemiol Public Heal* 2:1–25
- Eid D, Guzman-Rivero M, Rojas E et al (2018) Correction to: risk factors for cutaneous leishmaniasis in the rainforest of Bolivia: a cross-sectional study. *Trop Med Health*:46
- El Alem MMM, Hakkour M, Hmamouch A et al (2018) Risk factors and prediction analysis of cutaneous leishmaniasis due to *Leishmania tropica* in Southwestern Morocco. *Infect Genet Evol* 61:84–91. <https://doi.org/10.1016/j.meegid.2018.03.017>
- Elnaiem DEA, Mukhawi AM, Hassan MM et al (2003a) Factors affecting variations in exposure to infections by *Leishmania donovani* in eastern Sudan. *East Mediterr Health J* 9:827–836
- Elnaiem DEA, Schorscher J, Bendall A et al (2003b) Risk mapping of visceral leishmaniasis: The role of local variation in rainfall and altitude on the presence and incidence of kala-azar in eastern Sudan. *Am J Trop Med Hyg* 68:10–17
- Espejo JT, Le Pont F, Mouchet J et al (1989) Epidemiologie de la leishmaniose tegumentaire en Bolivie. 1. Description des zones d'étude et fréquence de la maladie. *Ann Soc Belg Med Trop* (1920) 69:297–306
- Fakhar M, Karamian M, Ghatee MA et al (2017) Distribution pattern of anthroponotic cutaneous leishmaniasis caused by *Leishmania tropica* in Western Afghanistan during 2013–2014. *Acta Trop* 176:22–28. <https://doi.org/10.1016/j.actatropica.2017.07.028>
- Faris R, Massoud A, El Said S et al (1988) The epidemiology of human visceral leishmaniasis in El Agamy (Alexandria Governorate), Egypt: serosurvey and case/control study. *Ann Trop Med Parasitol* 82:445–452. <https://doi.org/10.1080/00034983.1988.11812274>
- Faucher B, Gaudart J, Faraut F et al (2012) Heterogeneity of environments associated with transmission of visceral leishmaniasis in South-eastern France and implication for control strategies. *PLoS Negl Trop Dis* 6:1–9. <https://doi.org/10.1371/journal.pntd.0001765>
- Feliciangeli MD, Delgado O, Suarez B, Bravo A (2006) *Leishmania* and sand flies: Proximity to woodland as a risk factor for infection in a rural focus of visceral leishmaniasis in west central Venezuela. *Tropical Med Int Health* 11:1785–1791. <https://doi.org/10.1111/j.1365-3156.2006.01747.x>
- Ferro C, López M, Fuya P et al (2015) Spatial distribution of sand fly vectors and eco-epidemiology of cutaneous leishmaniasis transmission in Colombia. *PLoS One* 10:1–16. <https://doi.org/10.1371/journal.pone.0139391>
- Forattini OP, Rabello EX, Serra OP et al (1976) Observações sobre a transmissão da leishmaniose tegumentar no estado de São Paulo. *Brasil Rev Saude Publica*:31–43
- Galgamuwa LS, Dharmaratne SD, Iddawela D (2018) Leishmaniasis in Sri Lanka: spatial distribution and seasonal variations from 2009 to 2016. *Parasit Vectors* 11:60. <https://doi.org/10.1186/s13071-018-2647-5>
- Gao X, Cao Z (2019) Meteorological conditions, elevation and land cover as predictors for the distribution analysis of visceral leishmaniasis in

- Sinkiang province, Mainland China. *Sci Total Environ* 646:1111–1116. <https://doi.org/10.1016/j.scitotenv.2018.07.391>
- Gebremichael Tedla D, Bariagabr FH, Abreha HH (2018) Incidence and trends of Leishmaniasis and its risk factors in humera, Western Tigray. *J Parasitol Res*. <https://doi.org/10.1155/2018/8463097>
- Ghatee MA, Haghdoust AA, Kooreshnia F et al (2018) Role of environmental, climatic risk factors and livestock animals on the occurrence of cutaneous leishmaniasis in newly emerging focus in Iran. *J Infect Public Health* 11:425–433. <https://doi.org/10.1016/j.jiph.2017.12.004>
- Giannakopoulos A, Tsokana CN, Pervanidou D et al (2016) Environmental parameters as risk factors for human and canine Leishmania infection in Thessaly, Central Greece. *Parasitology* 143:1179–1186. <https://doi.org/10.1017/S0031182016000378>
- Gijón-Robles P, Abattouy N, Merino-Espinosa G et al (2018) Risk factors for the expansion of cutaneous leishmaniasis by Leishmania tropica: possible implications for control programmes. *Transbound Emerg Dis* 65:1615–1626. <https://doi.org/10.1111/tbed.12914>
- Gil JF, Nasser JR, Cajal SP et al (2010) Urban transmission of american cutaneous leishmaniasis in Argentina: spatial analysis study. *Am J Trop Med Hyg* 82:433–440. <https://doi.org/10.4269/ajtmh.2010.09-0113>
- Godana AA, Mwalili SM, Orwa GO (2019) Dynamic spatiotemporal modeling of the infected rate of visceral leishmaniasis in human in an endemic area of Amhara regional state, Ethiopia. *PLoS One* 14:e0212934. <https://doi.org/10.1371/journal.pone.0212934>
- Golpayegani AA, Moslem AR, Akhavan AA et al (2018) Modeling of environmental factors affecting the prevalence of zoonotic and anthroponotic cutaneous, and zoonotic visceral leishmaniasis in foci of Iran: a remote sensing and GIS based study. *J Arthropod Borne Dis* 12:41–66
- Gomez-Barroso D, Herrador Z, San Martin JV et al (2015) Spatial distribution and cluster analysis of a leishmaniasis outbreak in the South-western Madrid region, Spain, September 2009 to April 2013. *Euro Surveill* 20:1–10. <https://doi.org/10.2807/1560-7917.ES2015.20.7.21037>
- Grimaldi GJ, Tesh RB (1993) Leishmaniasis of the New World: current concepts and implications for future research. *Clin Microbiol Rev* 6:230–250. <https://doi.org/10.1128/CMR.6.3.230.Update>
- Guerra JADO, Guerra MDGB, Vasconcelos ZS et al (2019) Socioenvironmental aspects of the Purus Region - Brazilian Amazon: why relate them to the occurrence of American Tegumentary Leishmaniasis? *PLoS One* 14:e0211785. <https://doi.org/10.1371/journal.pone.0211785>
- Gutierrez JD, Martínez-Vega R, Ramoni-Perazzi J et al (2017) Environmental and socio-economic determinants associated with the occurrence of cutaneous leishmaniasis in the northeast of Colombia. *Trans R Soc Trop Med Hyg* 111:564–571. <https://doi.org/10.1093/trstmh/try011>
- Hasker E, Singh SP, Malaviya P et al (2012) Visceral leishmaniasis in rural Bihar, India. *Emerg Infect Dis* 18:1662–1664. <https://doi.org/10.3201/eid1810.111083>
- Haydon DT, Cleaveland S, Taylor LH, Laurenson MK (2002) Identifying reservoirs of infection: a conceptual and practical challenge. *Emerg Infect Dis* 8:1468–1473. <https://doi.org/10.3201/eid0812.010317>
- Hernández AM, Gutierrez JD, Xiao Y et al (2019) Spatial epidemiology of cutaneous leishmaniasis in Colombia: socioeconomic and demographic factors associated with a growing epidemic. *Trans R Soc Trop Med Hyg* 113:560–568. <https://doi.org/10.1093/trstmh/trz043>
- Holakouie-Naieni K, Mostafavi E, Boloorani AD et al (2017) Spatial modeling of cutaneous leishmaniasis in Iran from 1983 to 2013. *Acta Trop* 166:67–73. <https://doi.org/10.1016/j.actatropica.2016.11.004>
- Iddawela D, Vithana SMP, Atapattu D, Wijekoon L (2018) Clinical and epidemiological characteristics of cutaneous leishmaniasis in Sri Lanka. *BMC Infect Dis* 18:108. <https://doi.org/10.1186/s12879-018-2999-7>
- Iliopoulou P, Tsatsaris A, Katsios I et al (2018) Risk mapping of visceral leishmaniasis: a spatial regression model for Attica Region, Greece. *Trop Med Infect Dis* 3:83. <https://doi.org/10.3390/tropicalmed3030083>
- Jones TC, Johnson WD, Barretto AC et al (1987) Epidemiology of American cutaneous leishmaniasis due to *Leishmania braziliensis* brasiliensis. *J Infect Dis* 156:73–83
- Karagiannis-Voules DA, Scholte RGC, Guimarães LH et al (2013) Bayesian geostatistical modeling of leishmaniasis incidence in Brazil. *PLoS Negl Trop Dis* 7:13. <https://doi.org/10.1371/journal.pntd.0002213>
- Kariyawasam KKGDUL, Edirisuriya CS, Senerath U et al (2015) Characterisation of cutaneous leishmaniasis in Matara district, southern Sri Lanka: evidence for case clustering. *Pathog Glob Health* 109:1–8. <https://doi.org/10.1179/2047773215Y.0000000032>
- Kassem HA, Siri J, Kamal HA, Wilson ML (2012) Environmental factors underlying spatial patterns of sand flies (Diptera: Psychodidae) associated with leishmaniasis in southern Sinai, Egypt. *Acta Trop* 123:8–15. <https://doi.org/10.1016/j.actatropica.2012.02.067>
- Khanal B, Picado A, Bhattarai NR et al (2010) Spatial analysis of *Leishmania donovani* exposure in humans and domestic animals in a recent kala azar focus in Nepal. *Parasitology* 137:1597–1603. <https://doi.org/10.1017/S0031182010000521>
- Kirstein OD, Skrip L, Abassi I et al (2018) A fine scale eco-epidemiological study on endemic visceral leishmaniasis in north ethiopian villages. *Acta Trop* 183:64–77. <https://doi.org/10.1016/j.actatropica.2018.04.005>
- Kolaczinski JH, Reithinger R, Worku DT et al (2008) Risk factors of visceral leishmaniasis in East Africa: a case-control study in Pokot territory of Kenya and Uganda. *Int J Epidemiol* 37:344–352. <https://doi.org/10.1093/ije/dym275>
- Kumar N, Singh TB, Meena LP (2017) Socio-economic and behavioural risk factors of visceral leishmaniasis in the East Champaran district of Bihar. *Int J Community Med Public Health* 4:1577. <https://doi.org/10.18203/2394-6040.ijcmph20171767>
- Lainson R, Rangel BF (2005) *Lutzomyia longipalpis* and the eco-epidemiology of American visceral leishmaniasis, with particular reference to Brazil—a review. *Mem Inst Oswaldo Cruz* 100:811–827
- Lemma W, Tekie H, Yared S et al (2015) Sero-prevalence of *Leishmania donovani* infection in labour migrants and entomological risk factors in extra-domestic habitats of Kafta-Humera lowlands - kala-azar endemic areas in the northwest Ethiopia. *BMC Infect Dis* 15:1–8. <https://doi.org/10.1186/s12879-015-0830-2>
- Li Y, Zheng C (2019) Associations between meteorological factors and visceral leishmaniasis outbreaks in Jiashi County, Xinjiang Uygur Autonomous Region, China, 2005–2015. *Int J Environ Res Public Health* 16:1775. <https://doi.org/10.3390/ijerph16101775>
- Lima ID, Lima ALM, Mendes-Aguiar CO et al (2018) Changing demographics of visceral leishmaniasis in northeast Brazil: lessons for the future. *PLoS Negl Trop Dis* 12:e0006164. <https://doi.org/10.1371/journal.pntd.0006164>
- Loiseau R, Nabet C, Simon S et al (2019) American cutaneous leishmaniasis in French Guiana: an epidemiological update and study of environmental risk factors. *Int J Dermatol* 58:14625. <https://doi.org/10.1111/ijd.14625>
- López K, Tartaglino LC, Steinhorst II et al (2016) Factores de riesgo, representaciones y prácticas asociadas con la leishmaniasis visceral humana en un foco urbanoemergente en Posadas, Argentina. *Biomédica* 36:51–63
- Lopez J, Freire C, Moncayo T et al (2018) Prevalence and factors associated with cutaneous leishmaniasis in Tena-Napo, Ecuador 2012–2013. *Síndrome Cardiometaabólico y enfermedades crónica Degener* 8:5–9

- Lotukoi JA (2017) Exposure Factors Associated with Visceral Leishmaniasis (kala-azar) in Loima Sub-County of Turkana County, Kenya. *Am J Lab Med* 2:144. <https://doi.org/10.11648/j.ajlm.20170206.16>
- Machado-Coelho GLL, Assunção R, Mayrink W, Caiaffa WT (1999) American cutaneous leishmaniasis in Southeast Brazil: space-time clustering. *Int J Epidemiol* 28:982–989. <https://doi.org/10.1093/ije/28.5.982>
- Madalosso G, Fortaleza CM, Ribeiro AF et al (2012) American visceral leishmaniasis: factors associated with lethality in the state of São Paulo, Brazil. *J Trop Med* 2012:1–8. <https://doi.org/10.1155/2012/281572>
- Maia CS, Pimentel DS, Santana MA et al (2013) The perception of the risk factors associated with American visceral leishmaniasis in Petrolina, Pernambuco, Brazil. *Rev Med Vet (Bogota)* 7:19–25
- Maia Z, Viana V, Muniz E et al (2016) Risk factors associated with human visceral leishmaniasis in an urban area of Bahia, Brazil. *Vector-Borne Zoonotic Dis* 16:368–376. <https://doi.org/10.1089/vbz.2015.1880>
- Mandal P, Wagle RR, Thakur AK et al (2019) “Risk Factors for Visceral Leishmaniasis in Selected High Endemic Areas of Morang District, Nepal”: a case control Study. *bioRxiv*:530741. <https://doi.org/10.1101/530741>
- Melchior LAK, Brilhante AF, Chiaravalloti-Neto F (2017) Spatial and temporal distribution of American cutaneous leishmaniasis in Acre state, Brazil. *Infect Dis Poverty* 6:99. <https://doi.org/10.1186/s40249-017-0311-5>
- Melo HA, Rossoni DF, Teodoro U (2018) Effect of vegetation on cutaneous leishmaniasis in Paraná. *Brazil Mem Inst Oswaldo Cruz* 113. <https://doi.org/10.1590/0074-02760170505>
- Menezes JA, Luz TCB, de Sousa FF et al (2016) Fatores de risco peridomiciliares e conhecimento sobre leishmaniose visceral da população de Formiga, Minas Gerais. *Rev Bras Epidemiol* 19: 362–374. <https://doi.org/10.1590/1980-5497201600020013>
- Miranda C, Marques CCA, Massa JL (1998) Sensoriamento remoto orbital como recurso para análise da ocorrência da leishmaniose tegumentar americana em localidade urbana da região Sudeste do Brasil. *Rev Saude Publica* 32:455–463
- Mokhtari M, Miri M, Nikoonahad A et al (2016) Cutaneous leishmaniasis prevalence and morbidity based on environmental factors in Ilam, Iran: Spatial analysis and land use regression models. *Acta Trop* 163:90–97. <https://doi.org/10.1016/j.actatropica.2016.08.002>
- Monteiro WM, Neitzke-Abreu HC, Ferreira MEM et al (2009) Population mobility and production of American tegumentary leishmaniasis in the State of Paraná, southern Brazil. *Rev Soc Bras Med Trop* 42:509–514
- Montoya J, Jaramillo C, Palma G et al (1990) Report of an epidemic outbreak of tegumentary leishmaniasis in a coffee-growing area of Colombia. *Mem Inst Oswaldo Cruz* 85:119–121
- Moo Llanes DA (2016) Nicho ecológico actual y futuro de la leishmaniasis en la región Neotropical. *Rev Biol Trop* 64:1237–1245
- Moradiazl E, Rassi Y, Hanafi-Bojd AA et al (2018) The relationship between climatic factors and the prevalence of visceral leishmaniasis in North West of Iran. *Int. J Pediatr* 6:7169–7178. <https://doi.org/10.22038/ijp.2018.26837.2314>
- Moradi-Asl E, Hanafi-Bojd AA, Rassi Y et al (2017) Situational analysis of visceral leishmaniasis in the most important endemic area of the disease in Iran. *J Arthropod Borne Dis* 11:482–496
- Moreno EC, Melo MN, Genaro O et al (2005) Risk factors for Leishmania chagasi infection in an urban area of Minas Gerais State. *Rev Soc Bras Med Trop* 38:456–463. <https://doi.org/10.1590/S0037-86822005000600002>
- Moškovskij ŠD, Duhanina NN (1971) Epidemiology of the leishmaniasis: general considerations. *Bull World Health Organ* 44:529–534
- Mott KE, Desjeux P, Moncayo A et al (1990) Parasitic diseases and urban development. *Bull World Health Organ* 68:691–698
- Myers SS, Gaffikin L, Golden CD et al (2013) Human health impacts of ecosystem alteration. *Proc Natl Acad Sci U S A* 110:18753–18760. <https://doi.org/10.1073/pnas.1218656110>
- Nackers F, Mueller YK, Salih N et al (2015) Determinants of Visceral Leishmaniasis: a case-control study in Gedaref State, Sudan. *PLoS Negl Trop Dis* 9:1–16. <https://doi.org/10.1371/journal.pntd.0004187>
- Nasser JT, Donalisio MR, Vasconcelos CH (2009) Distribuição espacial dos casos de leishmaniose tegumentar americana no município de Campinas, Estado de São Paulo, no período de 1992 a 2003 Spatial distribution of American tegumentary leishmaniasis cases in Campinas, State of São Paulo, between 199. *Rev Soc Bras Med Trop* 42:309–314
- Navin TR, Sierra M, Custodio R et al (1985) Epidemiologic study of visceral leishmaniasis in Honduras, 1975–1983. *Am J Trop Med Hyg* 34:1069–1075
- Nazari M, Nazari S, Hanafi-Bojd AA et al (2017) Situation analysis of cutaneous leishmaniasis in an endemic area, south of Iran. *Asian Pac J Trop Med* 10:92–97. <https://doi.org/10.1016/j.apjtm.2016.12.001>
- Negera E, Gadisa E, Yamaiah L et al (2008) Outbreak of cutaneous leishmaniasis in Silti woreda, Ethiopia: risk factor assessment and causative agent identification. *Trans R Soc Trop Med Hyg* 102:883–890. <https://doi.org/10.1016/j.trstmh.2008.03.021>
- Nikonahad A, Khorshidi A, Ghaffari HR et al (2017) A time series analysis of environmental and metrological factors impact on cutaneous leishmaniasis incidence in an endemic area of Dehloran, Iran. *Environ Sci Pollut Res* 24:14117–14123. <https://doi.org/10.1007/s11356-017-8962-0>
- Nilfroushzadeh MA, Hosseini SM, Heidari A et al (2014) Domestic and peridomestic risk factors associated with transmission of cutaneous leishmaniasis in three hypo endemic, endemic, and hyper endemic areas: a randomized epidemiological study. *J Res Med Sci* 19:928–932
- Ocampo CB, Ferro MC, Cadena H et al (2012) Environmental factors associated with American cutaneous leishmaniasis in a new Andean focus in Colombia. *Tropical Med Int Health* 17:1309–1317. <https://doi.org/10.1111/j.1365-3156.2012.03065.x>
- Oliveira CDL, Diez-Roux A, César CC, Proietti FA (2006) A case-control study of microenvironmental risk factors for urban visceral leishmaniasis in a large city in Brazil, 1999–2000. *Pan Am J Public Health* 20:369–376. <https://doi.org/10.1590/S1020-49892006001100002>
- Oliveira AM, López RVM, Dibo MR et al (2018) Dispersion of Lutzomyia longipalpis and expansion of visceral leishmaniasis in São Paulo State, Brazil: identification of associated factors through survival analysis. *Parasit Vectors* 11:503. <https://doi.org/10.1186/s13071-018-3084-1>
- Pace D (2014) Leishmaniasis. *J Inf Secur* 20:1–9. <https://doi.org/10.1016/j.jinf.2014.07.016>
- Patz JA, Daszak P, Tabor GM et al (2004) Unhealthy landscapes: policy recommendations on land use change and infectious disease emergence. *Environ Health Perspect* 112:1092–1098. <https://doi.org/10.1289/ehp.6877>
- Pérez-Flórez M, Ocampo CB, Valderrama-Ardila C, Alexander N (2016) Spatial modeling of cutaneous leishmaniasis in the Andean region of Colombia. *Mem Inst Oswaldo Cruz* 111:433–442. <https://doi.org/10.1590/0074-02760160074>
- Perilla-González Y, Gómez-Suta D, Delgado-Osorio N et al (2014) Study of the scientific production on leishmaniasis in Latin America. *Recent Pat Antiinfect Drug Discov* 9:216–222
- Perry D, Dixon K, Garlapati R et al (2013) Visceral leishmaniasis prevalence and associated risk factors in the Saran District of Bihar, India, from 2009 to July of 2011. *Am J Trop Med Hyg* 88:778–784. <https://doi.org/10.4269/ajtmh.12-0442>
- Pigott DM, Bhatt S, Golding N et al (2014a) Global distribution maps of the leishmaniasis. *Elife*:1–21. <https://doi.org/10.7554/eLife.02851>

- Pigott DM, Golding N, Messina JP et al (2014b) Global database of leishmaniasis occurrence locations, 1960–2012. *Sci Data* 1:1–7. <https://doi.org/10.1038/sdata.2014.36>
- Ponte CB, Souza NC, Cavalcante MN et al (2011) Risk factors for *Leishmania chagasi* infection in an endemic area in Raposa, State of Maranhão, Brazil. *Rev Soc Bras Med Trop* 44:712–721. <https://doi.org/10.1590/S0037-86822011005000059>
- Prestes-Carneiro LE, Daniel LAF, Almeida LC et al (2019) Spatiotemporal analysis and environmental risk factors of visceral leishmaniasis in an urban setting in São Paulo State, Brazil. *Parasit Vectors* 12:251. <https://doi.org/10.1186/s13071-019-3496-6>
- Prudhomme J, Rahola N, Toty C et al (2015) Ecology and spatiotemporal dynamics of sandflies in the Mediterranean Languedoc region (Roquedur area, Gard, France). *Parasit Vectors* 8:1–14. <https://doi.org/10.1186/s13071-015-1250-2>
- Quintana MG, Fernández MS, Salomón OD (2012) Distribution and abundance of phlebotominae, vectors of leishmaniasis, in Argentina: Spatial and temporal analysis at different scales. *J Trop Med* 2012:1–16. <https://doi.org/10.1155/2012/652803>
- Rajabi M, Mansourian A, Pilsejo P, Bazmani A (2014) Environmental modelling of visceral leishmaniasis by susceptibility-mapping using neural networks: a case study in north-western Iran. *Geospat Health* 9:179–191. <https://doi.org/10.4081/gh.2014.15>
- Rajabi M, Pilesjö P, Bazmani A, Mansourian A (2016a) Identification of visceral leishmaniasis-susceptible areas using spatial modelling in Southern Caucasus. *Zoonoses Public Health*:1–18. <https://doi.org/10.1111/zph.12325>
- Rajabi M, Pilesjö P, Shirzadi MR et al (2016b) A spatially explicit agent-based modeling approach for the spread of cutaneous leishmaniasis disease in central Iran, Isfahan. *Environ Model Softw* 82:330–346. <https://doi.org/10.1016/j.envsoft.2016.04.006>
- Ramezankhani R, Hosseini A, Sajjadi N et al (2017a) Environmental risk factors for the incidence of cutaneous leishmaniasis in an endemic area of Iran: A GIS-based approach. *Spat Spatiotemporal Epidemiol*, Iran. <https://doi.org/10.1016/j.sste.2017.03.003>
- Ramezankhani R, Sajjadi N, Esmailzadeh RN et al (2017b) Spatial analysis of cutaneous leishmaniasis in an endemic area of Iran based on environmental factors. *Geospat Health* 12:282–293. <https://doi.org/10.4081/gh.2017.578>
- Ramezankhani R, Sajjadi N, Nezakati esmaeilzadeh R et al (2018) Climate and environmental factors affecting the incidence of cutaneous leishmaniasis in Isfahan, Iran. *Environ Sci Pollut Res* 25: 11516–11526. <https://doi.org/10.1007/s11356-018-1340-8>
- Ranasinghe S, Wickremasinghe R, Munasinghe A et al (2013) Cross-sectional study to assess risk factors for leishmaniasis in an endemic region in Sri Lanka. *Am J Trop Med Hyg* 89:742–749. <https://doi.org/10.4269/ajtmh.12-0640>
- Ranjan A, Sur D, Singh VP et al (2005) Risk factors for Indian kala-azar. *Am J Trop Med Hyg* 73:74–78
- Ready PD (2013) Biology of phlebotomine sand flies as vectors of disease agents. *Annu Rev Entomol* 58:227–250. <https://doi.org/10.1146/annurev-ento-120811-153557>
- Reithinger R, Mohsen M, Leslie T (2010) Risk factors for anthroponotic cutaneous leishmaniasis at the household level in Kabul, Afghanistan. *PLoS Negl Trop Dis* 4:1–8. <https://doi.org/10.1371/journal.pntd.0000639>
- Reyes A, Arrivillaga J (2009) Fauna Mammalia asociada a los focos de leishmaniasis neotropical: Situación en Venezuela. *Boletín Malariol y Salud Ambient* 49:35–52
- Rijal S, Urnaw S, Chappuis F et al (2010) Epidemiology of *Leishmania donovani* infection in high-transmission foci in Nepal. *Tropical Med Int Health* 15:21–28. <https://doi.org/10.1111/j.1365-3156.2010.02518.x>
- Rodrigues MGA, Sousa JDB, Dias ÁLB et al (2019) The role of deforestation on American cutaneous leishmaniasis incidence: spatial-temporal distribution, environmental and socioeconomic factors associated in the Brazilian Amazon. *Tropical Med Int Health* 24: 348–355. <https://doi.org/10.1111/tmi.13196>
- Rodríguez-Morales AJ, Pascual-González Y, Benítez JA et al (2010) Asociación entre la incidencia de leishmaniasis cutánea y el índice de desarrollo humano y sus componentes en cuatro estados endémicos de Venezuela. *Rev Peru Med Exp Salud Publica* 27: 22–30
- Roque ALR, Jansen AM (2014) Wild and synanthropic reservoirs of *Leishmania* species in the Americas. *Int J Parasitol Parasites Wildl* 3:251–262. <https://doi.org/10.1016/j.ijppaw.2014.08.004>
- Ruh E, Bostanci A, Kunter V et al (2017) Leishmaniasis in northern Cyprus: human cases and their association with risk factors. *J Vector Borne Dis* 54:358–365. <https://doi.org/10.4103/0972-9062.225842>
- Rutledge LC, Gupta RK (2002) Moth flies and sand flies (Psychodidae). In: *Medical and Veterinary Entomology*. Elsevier Science, pp 147–161
- Ryan JR, Mbui J, Rashid JR et al (2006) Spatial clustering and epidemiological aspects of visceral leishmaniasis in two endemic villages, Baringo District, Kenya. *Am J Trop Med Hyg* 74:308–317
- Salomón OD, Wilson ML, Munstermann LE, Travi BL (2004) Spatial and temporal patterns of phlebotomine sand flies (Diptera: Psychodidae) in a cutaneous leishmaniasis focus in Northern Argentina. *J Med Entomol* 41:33–39. <https://doi.org/10.1603/0022-2585-41.1.33>
- Salomón OD, Feliciangeli MD, Quintana MG et al (2015) *Lutzomyia longipalpis* urbanisation and control. *Mem Inst Oswaldo Cruz* 110: 831–846. <https://doi.org/10.1590/0074-02760150207>
- Schenkel K, Rijal S, Koirala S et al (2006) Visceral leishmaniasis in southeastern Nepal: a cross-sectional survey on *Leishmania donovani* infection and its risk factors. *Tropical Med Int Health* 11:1792–1799. <https://doi.org/10.1111/j.1365-3156.2006.01735.x>
- Seid A, Gadisa E, Tsegaw T et al (2014) Risk map for cutaneous leishmaniasis in Ethiopia based on environmental factors as revealed by geographical information systems and statistics. *Geospat Health* 8: 377–387. <https://doi.org/10.4081/gh.2014.27>
- Shah H (2005) Prevalence of visceral leishmaniasis associated with the behaviors of the people in rural areas. *J Nepal Med Assoc* 44:116–120
- Sharma U, Singh S (2008) Insect vectors of *Leishmania*: distribution, physiology and their control. *J Vector Borne Dis* 45:255–272
- Shaw J (2007) The leishmaniases—survival and expansion in a changing world. A mini-review. *Mem Inst Oswaldo Cruz* 102:541–546. <https://doi.org/10.1590/S0074-02762007000500001>
- Shiravand B, Tafti AAD, Hanafi-Bojd AA et al (2018) Modeling spatial risk of zoonotic cutaneous leishmaniasis in Central Iran. *Acta Trop* 185:327–335. <https://doi.org/10.1016/j.actatropica.2018.06.015>
- Singh NS, Singh PD (2019) A review on major risk factors and current status of visceral leishmaniasis in north India. *Am J Entomol* 3:6. <https://doi.org/10.11648/j.aje.20190301.12>
- Singh SP, Hasker E, Picado A et al (2010) Risk factors for visceral leishmaniasis in India: further evidence on the role of domestic animals. *Tropical Med Int Health* 15:29–35. <https://doi.org/10.1111/j.1365-3156.2010.02515.x>
- Sosa-Estani S, Segura EL, Gomez A et al (2001) Leishmaniose cutánea no Norte da Argentina: fatores de risco identificados num estudo caso-coorte em três municípios de Salta. *Rev Soc Bras Med Trop* 34:511–517
- Southgate BA (1964) Studies in the epidemiology of East African leishmaniasis. 2. The human distribution and its determinants. *Trans R Soc Trop Med Hyg* 58:377–390
- Souza VAF, Cortez LR, Dias RA et al (2012) Space-time cluster analysis of American visceral leishmaniasis in Bauru, São Paulo State, Brazil. *Cad Saude Publica* 28:1949–1964. <https://doi.org/10.1590/S0102-311X2012001000013>

- Steffens I (2010) Vector-borne diseases Special edition. *Eur J Infect*:1–124
- Talmoudi K, Bellali H, Ben-Alaya N et al (2017) Modeling zoonotic cutaneous leishmaniasis incidence in central Tunisia from 2009–2015: Forecasting models using climate variables as predictors. *PLoS Negl Trop Dis* 11:1–18. <https://doi.org/10.1371/journal.pntd.0005844>
- Temponi AOD, de Brito MG, Ferraz ML et al (2018) Ocorrência de casos de leishmaniose tegumentar americana: uma análise multivariada dos circuitos espaciais de produção, Minas Gerais, Brasil, 2007 a 2011. *Cad Saude Publica* 34:1–15. <https://doi.org/10.1590/0102-311x00165716>
- Thompson RA, Wellington de Oliveira Lima J, Maguire JH et al (2002) Climatic and demographic determinants of American visceral leishmaniasis in northeastern Brazil using remote sensing technology for environmental categorization of rain and region influences on leishmaniasis. *Am J Trop* 67:648–655
- Toledo CRS, Almeida AS, Chaves SAM et al (2017) Vulnerability to the transmission of human visceral leishmaniasis in a Brazilian urban area. *Rev Saude Publica* 51:49. <https://doi.org/10.1590/S1518-8787.2017051006532>
- Tsegaw T, Gadisa E, Seid A et al (2013) Identification of environmental parameters and risk mapping of visceral leishmaniasis in Ethiopia by using geographical information systems and a statistical approach. *Geospat Health* 7:299–308. <https://doi.org/10.4081/gh.2013.88>
- Ullah K, Khan NH, Sepúlveda N et al (2016) Assessing incidence patterns and risk factors for cutaneous leishmaniasis in Peshawar Region, Khyber Pakhtunkhwa, Pakistan. *J Parasitol* 102:501–506. <https://doi.org/10.1645/15-919>
- Uranw S, Hasker E, Roy L et al (2013) An outbreak investigation of visceral leishmaniasis among residents of Dharan town, Eastern Nepal, evidence for urban transmission of *Leishmania donovani*. *BMC Infect Dis* 13:9. <https://doi.org/10.1186/1471-2334-13-21>
- Valderrama-Ardila C, Alexander N, Ferro C et al (2010) Environmental risk factors for the incidence of American cutaneous leishmaniasis in a sub-andean zone of Colombia (Chaparral, Tolima). *Am J Trop Med Hyg* 82:243–250. <https://doi.org/10.4269/ajtmh.2010.09-0218>
- Viana GM, Nascimento Mdo D, Rabelo EM et al (2011) Relationship between rainfall and temperature: observations on the cases of visceral leishmaniasis in São Luis Island, State of Maranhão, Brazil. *Rev Soc Bras Med Trop* 44:722–724. <https://doi.org/10.1590/S0037-86822011000600013>
- Votýpka J, Kasap OE, Volf P et al (2012) Risk factors for cutaneous leishmaniasis in Cukurova region, Turkey. *Trans R Soc Trop Med Hyg* 106:186–190. <https://doi.org/10.1016/j.trstmh.2011.12.004>
- Wade TG, Riitters KH, Wickham JD, Jones KB (2003) Distribution and causes of global forest fragmentation. *Ecol Soc* 7
- Weerakoon HS, Ranawaka RR, Warnasekara J, Bandara P (2018) Analysis of gap in the correct diagnosis of leishmaniasis at primary care level, socio demographic and environmental risk factors of leishmaniasis transmission in Anuradhapura. In: Anuradhapura Med J
- Weigle K, Santrich C, Martinez F et al (1993) Epidemiology of cutaneous leishmaniasis in Colombia: environmental and behavioral risk factors for infection, clinical manifestations, and pathogenicity. *J Infect Dis* 168:709–714
- Werneck GL, Costa CHN, Walker AM et al (2002) The urban spread of visceral leishmaniasis: clues from spatial analysis. *Epidemiology* 13:364–367. <https://doi.org/10.1097/00001648-200205000-00020>
- Werneck GL, Costa CHN, Walker AM et al (2007) Multilevel modelling of the incidence of visceral leishmaniasis in Teresina, Brazil. *Epidemiol Infect* 135:195–201. <https://doi.org/10.1017/S0950268806006881>
- Wood CL, Lafferty KD, DeLeo G et al (2014) Does biodiversity protect humans against infectious disease? *Ecology* 95:817–832. <https://doi.org/10.1890/13-1041.1>
- World Health Organization (2010) Control of the leishmaniases. *World Health Organ Tech Rep Ser*:1–202. <https://doi.org/10.1038/nmicro1766>
- World Health Organization (2017) Global leishmaniasis update, 2006–2015: a turning point in leishmaniasis surveillance. *World Heal Organ Geneva* 92:557–572. <https://doi.org/10.1186/1750-9378-2-15.Voir>
- World Health Organization (2018) Global Health Observatory (GHO) data| Leishmaniasis. In: WHO. https://www.who.int/gho/neglected_diseases/leishmaniasis/en/. Accessed 2 Mar 2019
- Yadon ZE, Rodrigues LC, Davies CR, Quigley MA (2003) Indoor and peridomestic transmission of American cutaneous leishmaniasis in northwestern Argentina: a retrospective case-control study. *Am J Trop Med Hyg* 68:519–526. <https://doi.org/10.4269/ajtmh.2003.68.519>
- Yared S, Deribe K, Gebreselassie A et al (2014) Risk factors of visceral leishmaniasis: a case control study in north-western Ethiopia. *Parasit Vectors* 7:1–11. <https://doi.org/10.1186/s13071-014-0470-1>
- Zheng C, Fu J, Li Z et al (2018) Spatiotemporal variation and hot spot detection of visceral leishmaniasis disease in Kashi Prefecture, China. *Int J Environ Res Public Health* 15:2784. <https://doi.org/10.3390/ijerph15122784>
- Zorrilla V, Agüero M, Cáceres A et al (2005) Factores de riesgo que determinan la transmisión de la leishmaniasis en el valle Llaucano, Chota-Cajamarca. *Info* 42:33–42

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